

The Enzyme List

Class 5 — Isomerases

Nomenclature Committee
of the
International Union of Biochemistry and Molecular Biology
(NC-IUBMB)

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EC 5.1 Racemases and epimerases

This subclass contains enzymes that catalyse either racemization or epimerization of a centre of chirality. Sub-subclasses are based on the substrate: amino acids and derivatives (EC 5.1.1), hydroxy acids and derivatives (EC 5.1.2), carbohydrates and derivatives (EC 5.1.3), or other compounds (EC 5.1.99).

EC 5.1.1 Acting on amino acids and derivatives

EC 5.1.1.1

Accepted name: alanine racemase
Reaction: L-alanine = D-alanine
Other name(s): L-alanine racemase
Systematic name: alanine racemase
Comments: A pyridoxal-phosphate protein.
References: [176, 323, 324]

[EC 5.1.1.1 created 1961]

EC 5.1.1.2

Accepted name: methionine racemase
Reaction: L-methionine = D-methionine
Systematic name: methionine racemase
Comments: A pyridoxal-phosphate protein.
References: [131]

[EC 5.1.1.2 created 1961]

EC 5.1.1.3

Accepted name: glutamate racemase
Reaction: L-glutamate = D-glutamate
Systematic name: glutamate racemase
Comments: A pyridoxal-phosphate protein.
References: [94]

[EC 5.1.1.3 created 1961]

EC 5.1.1.4

Accepted name: proline racemase
Reaction: L-proline = D-proline
Systematic name: proline racemase
References: [269]

[EC 5.1.1.4 created 1961]

EC 5.1.1.5

Accepted name: lysine racemase
Reaction: L-lysine = D-lysine
Systematic name: lysine racemase
References: [118]

[EC 5.1.1.5 created 1961]

EC 5.1.1.6

Accepted name: threonine racemase
Reaction: L-threonine = D-threonine
Systematic name: threonine racemase
Comments: Inverts both chiral centres.
References: [9]

[EC 5.1.1.6 created 1961, modified 1981]

EC 5.1.1.7

Accepted name: diaminopimelate epimerase
Reaction: LL-2,6-diaminoheptanedioate = *meso*-diaminoheptanedioate
Systematic name: LL-2,6-diaminoheptanedioate 2-epimerase
References: [14]

[EC 5.1.1.7 created 1961]

EC 5.1.1.8

Accepted name: 4-hydroxyproline epimerase
Reaction: *trans*-4-hydroxy-L-proline = *cis*-4-hydroxy-D-proline
Other name(s): hydroxyproline epimerase; hydroxyproline 2-epimerase; L-hydroxyproline epimerase
Systematic name: 4-hydroxyproline 2-epimerase
Comments: Also interconverts *trans*-4-hydroxy-D-proline and *cis*-4-hydroxy-L-proline.
References: [2]

[EC 5.1.1.8 created 1965, modified 1983]

EC 5.1.1.9

Accepted name: arginine racemase
Reaction: L-arginine = D-arginine
Systematic name: arginine racemase
Comments: A pyridoxal-phosphate protein.
References: [335]

[EC 5.1.1.9 created 1972]

EC 5.1.1.10

Accepted name: amino-acid racemase
Reaction: an L-amino acid = a D-amino acid
Other name(s): L-amino acid racemase
Systematic name: amino-acid racemase
Comments: A pyridoxal-phosphate protein.
References: [265]

[EC 5.1.1.10 created 1972]

EC 5.1.1.11

Accepted name: phenylalanine racemase (ATP-hydrolysing)
Reaction: ATP + L-phenylalanine + H₂O = AMP + diphosphate + D-phenylalanine
Other name(s): phenylalanine racemase; phenylalanine racemase (adenosine triphosphate-hydrolysing); gramicidin S synthetase I
Systematic name: phenylalanine racemase (ATP-hydrolysing)
References: [329]

[EC 5.1.1.11 created 1972]

EC 5.1.1.12

Accepted name: ornithine racemase
Reaction: L-ornithine = D-ornithine

Systematic name: ornithine racemase
References: [47]

[EC 5.1.1.12 created 1972 as EC 5.4.3.1, transferred 1976 to EC 5.1.1.12]

EC 5.1.1.13

Accepted name: aspartate racemase
Reaction: L-aspartate = D-aspartate
Other name(s): D-aspartate racemase; McyF
Systematic name: aspartate racemase
Comments: Also acts, at half the rate, on L-alanine.
References: [160, 332, 168, 259, 331]

[EC 5.1.1.13 created 1976]

EC 5.1.1.14

Accepted name: nocardicin-A epimerase
Reaction: isonocardicin A = nocardicin A
Other name(s): isonocardicin A epimerase
Systematic name: nocardicin-A epimerase
Comments: The (9'S) configuration of isonocardicin A is converted into the (9'R) configuration.
References: [313]

[EC 5.1.1.14 created 1992]

EC 5.1.1.15

Accepted name: 2-aminohexano-6-lactam racemase
Reaction: L-2-aminohexano-6-lactam = D-2-aminohexano-6-lactam
Other name(s): α -amino- ϵ -caprolactam racemase
Systematic name: 2-aminohexano-6-lactam racemase
Comments: Contains pyridoxal 5'-phosphate. Also racemises 2-aminopentano-5-lactam (α -amino- δ -valerolactam) and 2-amino-4-thiahexano-6-lactam (where S replaces CH₂ of C-4). It does not catalyse the racemisation of α -amino acids but has some transaminase activity with them.
References: [4, 5]

[EC 5.1.1.15 created 1999]

EC 5.1.1.16

Accepted name: protein-serine epimerase
Reaction: [protein]-L-serine = [protein]-D-serine
Other name(s): protein-serine racemase
Systematic name: [protein]-serine epimerase
Comments: The enzyme specifically interconverts the configuration of Ser-46 of the peptide ω -agatoxin-KT, found in the venom of the funnel web spider, *Agelenopsis aperta*, but not that of the other serine residue, Ser-28.
References: [256]

[EC 5.1.1.16 created 1999]

EC 5.1.1.17

Accepted name: isopenicillin-N epimerase
Reaction: isopenicillin N = penicillin N

Systematic name: penicillin-N 5-amino-5-carboxypentanoyl-epimerase
Comments: This enzyme contains pyridoxal phosphate. Epimerization at C-5 of the 5-amino-5-carboxypentanoyl group to form penicillin N is required to make a substrate for EC 1.14.20.1, deacetoxycephalosporin-C synthase, to produce cephalosporins. Forms part of the penicillin biosynthesis pathway (for pathway, [click here](#)).
References: [\[299, 159, 43, 333\]](#)

[EC 5.1.1.17 created 2002]

EC 5.1.1.18

Accepted name: serine racemase
Reaction: L-serine = D-serine
Other name(s): SRR
Systematic name: serine racemase
Comments: A pyridoxal-phosphate protein that is highly selective for L-serine as substrate. D-Serine is found in type-II astrocytes in mammalian brain, where it appears to be an endogenous ligand of the glycine site of *N*-methyl-D-aspartate (NMDA) receptors [\[321, 322\]](#). The reaction can also occur in the reverse direction but does so more slowly at physiological serine concentrations [\[82\]](#).
References: [\[321, 322, 196, 82\]](#)

[EC 5.1.1.18 created 2007]

EC 5.1.2 Acting on hydroxy acids and derivatives

EC 5.1.2.1

Accepted name: lactate racemase
Reaction: (*S*)-lactate = (*R*)-lactate
Other name(s): lacticoracemase; hydroxyacid racemase; lactic acid racemase
Systematic name: lactate racemase
References: [\[120, 145\]](#)

[EC 5.1.2.1 created 1961]

EC 5.1.2.2

Accepted name: mandelate racemase
Reaction: (*S*)-mandelate = (*R*)-mandelate
Systematic name: mandelate racemase
References: [\[101\]](#)

[EC 5.1.2.2 created 1961]

EC 5.1.2.3

Accepted name: 3-hydroxybutyryl-CoA epimerase
Reaction: (*S*)-3-hydroxybutanoyl-CoA = (*R*)-3-hydroxybutanoyl-CoA
Other name(s): 3-hydroxybutyryl coenzyme A epimerase; 3-hydroxyacyl-CoA epimerase
Systematic name: 3-hydroxybutanoyl-CoA 3-epimerase
References: [\[272, 307\]](#)

[EC 5.1.2.3 created 1961]

EC 5.1.2.4

Accepted name: acetoin racemase
Reaction: (*S*)-acetoin = (*R*)-acetoin
Other name(s): acetylmethylcarbinol racemase
Systematic name: acetoin racemase
References: [284]

[EC 5.1.2.4 created 1972]

EC 5.1.2.5

Accepted name: tartrate epimerase
Reaction: (*R,R*)-tartrate = *meso*-tartrate
Other name(s): tartaric racemase
Systematic name: tartrate epimerase
References: [232]

[EC 5.1.2.5 created 1972]

EC 5.1.2.6

Accepted name: isocitrate epimerase
Reaction: (*1R,2S*)-1-hydroxypropane-1,2,3-tricarboxylate = (*1S,2S*)-1-hydroxypropane-1,2,3-tricarboxylate
Systematic name: (*1R,2S*)-1-hydroxypropane-1,2,3-tricarboxylate 1-epimerase
Comments: (*1R,2S*)-1-hydroxypropane-1,2,3-tricarboxylate is the commonly occurring isomer of isocitrate.
References: [116]

[EC 5.1.2.6 created 1984]

EC 5.1.3 Acting on carbohydrates and derivatives

EC 5.1.3.1

Accepted name: ribulose-phosphate 3-epimerase
Reaction: D-ribulose 5-phosphate = D-xylulose 5-phosphate
Other name(s): phosphoribulose epimerase; erythrose-4-phosphate isomerase; phosphoketopentose 3-epimerase; xy-lulose phosphate 3-epimerase; phosphoketopentose epimerase; ribulose 5-phosphate 3-epimerase; D-ribulose phosphate-3-epimerase; D-ribulose 5-phosphate epimerase; D-ribulose-5-P 3-epimerase; D-xylulose-5-phosphate 3-epimerase; pentose-5-phosphate 3-epimerase
Systematic name: D-ribulose-5-phosphate 3-epimerase
Comments: The enzyme also converts D-erythrose 4-phosphate into D-erythrulose 4-phosphate and D-threose 4-phosphate.
References: [16, 64, 121, 276, 286]

[EC 5.1.3.1 created 1961, modified 1989]

EC 5.1.3.2

Accepted name: UDP-glucose 4-epimerase
Reaction: UDP-glucose = UDP-galactose
Other name(s): UDP-galactose 4-epimerase; uridine diphosphoglucose epimerase; galactowaldenase; UDPG-4-epimerase; uridine diphosphate galactose 4-epimerase; uridine diphospho-galactose-4-epimerase; UDP-glucose epimerase; UDP-galactose 4-epimerase; 4-epimerase; UDPG-4-epimerase; uridine diphosphoglucose 4-epimerase; uridine diphosphate glucose 4-epimerase; UDP-D-galactose 4-epimerase
Systematic name: UDP-glucose 4-epimerase
Comments: Requires NAD⁺. Also acts on UDP-2-deoxyglucose.
References: [164, 186, 314]

[EC 5.1.3.2 created 1961]

EC 5.1.3.3

Accepted name: aldose 1-epimerase
Reaction: α -D-glucose = β -D-glucose
Other name(s): mutarotase; aldose mutarotase; galactose mutarotase; galactose 1-epimerase; D-galactose 1-epimerase
Systematic name: aldose 1-epimerase
Comments: Also acts on L-arabinose, D-xylose, D-galactose, maltose and lactose. This enzyme catalyses the first step in galactose metabolism by converting β -D-glucose into α -D-glucose, which is the substrate for EC 2.7.1.6, galactokinase [24, 289].
References: [27, 28, 140, 166, 24, 289, 288]

[EC 5.1.3.3 created 1961]

EC 5.1.3.4

Accepted name: L-ribulose-5-phosphate 4-epimerase
Reaction: L-ribulose 5-phosphate = D-xylulose 5-phosphate
Other name(s): phosphoribulose isomerase; ribulose phosphate 4-epimerase; L-ribulose-phosphate 4-epimerase; L-ribulose 5-phosphate 4-epimerase; AraD; L-Ru5P
Systematic name: L-ribulose-5-phosphate 4-epimerase
Comments: Requires a divalent cation for activity.
References: [42, 62, 162, 320, 12, 161, 245]

[EC 5.1.3.4 created 1965, modified 2005]

EC 5.1.3.5

Accepted name: UDP-arabinose 4-epimerase
Reaction: UDP-L-arabinose = UDP-D-xylose
Other name(s): uridine diphosphoarabinose epimerase; UDP arabinose epimerase; uridine 5'-diphosphate-D-xylose 4-epimerase; UDP-D-xylose 4-epimerase
Systematic name: UDP-L-arabinose 4-epimerase
References: [74]

[EC 5.1.3.5 created 1965]

EC 5.1.3.6

Accepted name: UDP-glucuronate 4-epimerase
Reaction: UDP-glucuronate = UDP-D-galacturonate
Other name(s): uridine diphospho-D-galacturonic acid; UDP glucuronic epimerase; uridine diphosphoglucuronic epimerase; UDP-galacturonate 4-epimerase; uridine diphosphoglucuronate epimerase; UDP-D-galacturonic acid 4-epimerase
Systematic name: UDP-glucuronate 4-epimerase
References: [74]

[EC 5.1.3.6 created 1965]

EC 5.1.3.7

Accepted name: UDP-N-acetylglucosamine 4-epimerase
Reaction: UDP-N-acetyl-D-glucosamine = UDP-N-acetyl-D-galactosamine
Other name(s): UDP acetylglucosamine epimerase; uridine diphosphoacetylglucosamine epimerase; uridine diphosphate N-acetylglucosamine-4-epimerase; uridine 5'-diphospho-N-acetylglucosamine-4-epimerase
Systematic name: UDP-N-acetyl-D-glucosamine 4-epimerase
References: [93, 153]

[EC 5.1.3.7 created 1965]

EC 5.1.3.8

Accepted name: *N*-acylglucosamine 2-epimerase
Reaction: *N*-acyl-D-glucosamine = *N*-acyl-D-mannosamine
Other name(s): acylglucosamine 2-epimerase; *N*-acetylglucosamine 2-epimerase
Systematic name: *N*-acyl-D-glucosamine 2-epimerase
Comments: Requires catalytic amounts of ATP.
References: [92]

[EC 5.1.3.8 created 1972]

EC 5.1.3.9

Accepted name: *N*-acylglucosamine-6-phosphate 2-epimerase
Reaction: *N*-acyl-D-glucosamine 6-phosphate = *N*-acyl-D-mannosamine 6-phosphate
Other name(s): acylglucosamine-6-phosphate 2-epimerase; acylglucosamine phosphate 2-epimerase
Systematic name: *N*-acyl-D-glucosamine-6-phosphate 2-epimerase
References: [91]

[EC 5.1.3.9 created 1972]

EC 5.1.3.10

Accepted name: CDP-paratose 2-epimerase
Reaction: CDP-3,6-dideoxy-D-glucose = CDP-3,6-dideoxy-D-mannose
Other name(s): CDP-paratose epimerase; cytidine diphosphoabequose epimerase; cytidine diphosphodideoxyglucose epimerase; cytidine diphosphoparatose epimerase; cytidine diphosphate paratose-2-epimerase; CDP-abequose epimerase (incorrect); CDP-D-abequose 2-epimerase (incorrect)
Systematic name: CDP-3,6-dideoxy-D-glucose 2-epimerase
Comments: Requires NAD⁺. CDP-paratose (CDP-3,6-dideoxy-D-glucose), is more systematically called CDP- α -3,6-dideoxy-D-*ribo*-hexose. CDP-tyvelose (CDP-3,6-dideoxy-D-mannose) is systematically called CDP-3,6-dideoxy-D-*arabino*-hexose.
References: [185, 167]

[EC 5.1.3.10 created 1972, modified 2005]

EC 5.1.3.11

Accepted name: cellobiose epimerase
Reaction: cellobiose = D-glucosyl-D-mannose
Systematic name: cellobiose 2-epimerase
References: [296]

[EC 5.1.3.11 created 1972]

EC 5.1.3.12

Accepted name: UDP-glucuronate 5'-epimerase
Reaction: UDP-glucuronate = UDP-L-iduronate
Other name(s): uridine diphosphoglucuronate 5'-epimerase; UDP-glucuronic acid 5'-epimerase; C-5-uronosyl epimerase
Systematic name: UDP-glucuronate 5'-epimerase
Comments: Requires NAD⁺.
References: [125]

[EC 5.1.3.12 created 1972]

EC 5.1.3.13

Accepted name: dTDP-4-dehydrorhamnose 3,5-epimerase
Reaction: dTDP-4-dehydro-6-deoxy-D-glucose = dTDP-4-dehydro-6-deoxy-L-mannose
Other name(s): dTDP-L-rhamnose synthetase; dTDP-L-rhamnose synthetase; thymidine diphospho-4-ketorhamnose 3,5-epimerase; TDP-4-ketorhamnose 3,5-epimerase; dTDP-4-dehydro-6-deoxy-D-glucose 3,5-epimerase; TDP-4-keto-L-rhamnose-3,5-epimerase
Systematic name: dTDP-4-dehydro-6-deoxy-D-glucose 3,5-epimerase
Comments: The enzyme occurs in a complex with EC 1.1.1.133 dTDP-4-dehydrorhamnose reductase.
References: [88, 189]

[EC 5.1.3.13 created 1972]

EC 5.1.3.14

Accepted name: UDP-*N*-acetylglucosamine 2-epimerase
Reaction: UDP-*N*-acetyl-D-glucosamine = UDP-*N*-acetyl-D-mannosamine
Other name(s): UDP-*N*-acetylglucosamine 2'-epimerase; uridine diphosphoacetylglucosamine 2'-epimerase; uridine diphospho-*N*-acetylglucosamine 2'-epimerase; uridine diphosphate-*N*-acetylglucosamine-2'-epimerase
Systematic name: UDP-*N*-acetyl-D-glucosamine 2-epimerase
Comments: The enzyme hydrolyses the product to UDP and *N*-acetyl-D-mannosamine.
References: [142]

[EC 5.1.3.14 created 1976]

EC 5.1.3.15

Accepted name: glucose-6-phosphate 1-epimerase
Reaction: α -D-glucose 6-phosphate = β -D-glucose 6-phosphate
Systematic name: D-glucose-6-phosphate 1-epimerase
References: [326]

[EC 5.1.3.15 created 1976]

EC 5.1.3.16

Accepted name: UDP-glucosamine 4-epimerase
Reaction: UDP-glucosamine = UDP-galactosamine
Systematic name: UDP-glucosamine 4-epimerase
References: [173, 260]

[EC 5.1.3.16 created 1984]

EC 5.1.3.17

Accepted name: heparosan-*N*-sulfate-glucuronate 5-epimerase
Reaction: heparosan-*N*-sulfate D-glucuronate = heparosan-*N*-sulfate L-iduronate
Other name(s): heparosan epimerase; heparosan-*N*-sulfate-D-glucuronosyl 5-epimerase; C-5 uronosyl epimerase; polyglucuronate epimerase; D-glucuronosyl C-5 epimerase; poly[(1,4)- β -D-glucuronosyl-(1,4)-*N*-sulfo- α -D-glucosaminyl] glucurono-5-epimerase
Systematic name: poly[(1 \rightarrow 4)- β -D-glucuronosyl-(1 \rightarrow 4)-*N*-sulfo- α -D-glucosaminyl] glucurono-5-epimerase
Comments: Acts on D-glucuronosyl residues adjacent to sulfated D-glucosamine units in the heparin precursor. Not identical with EC 5.1.3.19 chondroitin-glucuronate 5-epimerase.
References: [126]

[EC 5.1.3.17 created 1984]

EC 5.1.3.18

Accepted name: GDP-mannose 3,5-epimerase
Reaction: GDP-mannose = GDP-L-galactose
Other name(s): GDP-D-mannose:GDP-L-galactose epimerase; guanosine 5'-diphosphate D-mannose:guanosine 5'-diphosphate L-galactose epimerase
Systematic name: GDP-mannose 3,5-epimerase
References: [19, 110]

[EC 5.1.3.18 created 1986]

EC 5.1.3.19

Accepted name: chondroitin-glucuronate 5-epimerase
Reaction: chondroitin D-glucuronate = dermatan L-iduronate
Other name(s): polyglucuronate 5-epimerase; dermatan-sulfate 5-epimerase; uronosyl C-5 epimerase; chondroitin D-glucuronosyl 5-epimerase
Systematic name: chondroitin-D-glucuronate 5-epimerase
Comments: Not identical with EC 5.1.3.17 heparosan-*N*-sulfate-glucuronate 5-epimerase.
References: [174]

[EC 5.1.3.19 created 1986]

EC 5.1.3.20

Accepted name: ADP-*glyceromanno*-heptose 6-epimerase
Reaction: ADP-D-*glycero*-D-*manno*-heptose = ADP-L-*glycero*-D-*manno*-heptose
Systematic name: ADP-L-*glycero*-D-*manno*-heptose 6-epimerase
Comments: Requires NAD⁺.
References: [65, 229]

[EC 5.1.3.20 created 1999]

EC 5.1.3.21

Accepted name: maltose epimerase
Reaction: α -maltose = β -maltose
Systematic name: maltose 1-epimerase
Comments: The enzyme catalyses the interconversion of α and β anomers of maltose more effectively than those of disaccharides such as lactose and cellobiose.
References: [258]

[EC 5.1.3.21 created 2002]

EC 5.1.3.22

Accepted name: L-ribulose-5-phosphate 3-epimerase
Reaction: L-ribulose 5-phosphate = L-xylulose 5-phosphate
Other name(s): L-xylulose 5-phosphate 3-epimerase; UlaE; SgaU
Systematic name: L-ribulose-5-phosphate 3-epimerase
Comments: Along with EC 4.1.1.85, 3-dehydro-L-gulonate-6-phosphate decarboxylase, this enzyme is involved in a pathway for the utilization of L-ascorbate by *Escherichia coli*.
References: [334]

[EC 5.1.3.22 created 2005]

EC 5.1.3.23

- Accepted name:** UDP-2,3-diacetamido-2,3-dideoxyglucuronic acid 2-epimerase
Reaction: UDP-2,3-diacetamido-2,3-dideoxy- α -D-glucuronate = UDP-2,3-diacetamido-2,3-dideoxy- α -D-mannuronate
Other name(s): UDP-GlcNAc3NAcA 2-epimerase; UDP- α -D-GlcNAc3NAcA 2-epimerase; 2,3-diacetamido-2,3-dideoxy- α -D-glucuronic acid 2-epimerase; WbpI; WlbD
Systematic name: 2,3-diacetamido-2,3-dideoxy- α -D-glucuronate 2-epimerase
Comments: This enzyme participates in the biosynthetic pathway for UDP- α -D-ManNAc3NAcA (UDP-2,3-diacetamido-2,3-dideoxy- α -D-mannuronic acid), an important precursor of the B-band lipopolysaccharide of *Pseudomonas aeruginosa* serotype O5 and of the band-A trisaccharide of *Bordetella pertussis*, both important respiratory pathogens [311]. The enzyme is highly specific as UDP- α -D-GlcNAc, UDP- α -D-GlcNAcA (UDP-2-acetamido-2-deoxy- α -D-glucuronic acid) and UDP- α -D-GlcNAc3NAc (UDP-2,3-diacetamido-2,3-dideoxy- α -D-glucose) cannot act as substrates [311].
References: [311, 310, 135]

[EC 5.1.3.23 created 2007]

EC 5.1.99 Acting on other compounds

EC 5.1.99.1

- Accepted name:** methylmalonyl-CoA epimerase
Reaction: (*R*)-methylmalonyl-CoA = (*S*)-methylmalonyl-CoA
Other name(s): methylmalonyl-CoA racemase; methylmalonyl coenzyme A racemase; DL-methylmalonyl-CoA racemase; 2-methyl-3-oxopropanoyl-CoA 2-epimerase [incorrect]
Systematic name: methylmalonyl-CoA 2-epimerase
References: [187, 218]

[EC 5.1.99.1 created 1965, modified 1981]

EC 5.1.99.2

- Accepted name:** 16-hydroxysteroid epimerase
Reaction: 16 α -hydroxysteroid = 16 β -hydroxysteroid
Systematic name: 16-hydroxysteroid 16-epimerase
References: [56]

[EC 5.1.99.2 created 1972]

EC 5.1.99.3

- Accepted name:** allantoin racemase
Reaction: (*S*)(+)-allantoin = (*R*)(-)-allantoin
Systematic name: allantoin racemase
References: [302]

[EC 5.1.99.3 created 1976]

EC 5.1.99.4

- Accepted name:** α -methylacyl-CoA racemase
Reaction: (2*S*)-2-methylacyl-CoA = (2*R*)-2-methylacyl-CoA
Systematic name: 2-methylacyl-CoA 2-epimerase
Comments: α -methyl-branched acyl-CoA derivatives with chain lengths of more than C10 are substrates. Also active towards some aromatic compounds (e.g. ibuprofen) and bile acid intermediates, such as trihydroxycoprostanoyl-CoA. Not active towards free acids
References: [248]

[EC 5.1.99.4 created 1999]

EC 5.1.99.5

Accepted name: hydantoin racemase
Reaction: D-5-monosubstituted hydantoin = L-5-monosubstituted hydantoin
Other name(s): 5'-monosubstituted-hydantoin racemase; HyuA; HyuE
Systematic name: D-5-monosubstituted-hydantoin racemase
Comments: This enzyme, along with *N*-carbamoylase (EC 3.5.1.77 and EC 3.5.1.87) and hydantoinase, forms part of the reaction cascade known as the "hydantoinase process", which allows the total conversion of D,L-5-monosubstituted hydantoins into optically pure D- or L-amino acids [8]. The enzyme from *Pseudomonas* sp. (HyuE) has a preference for hydantoins with aliphatic substituents, e.g. D- and L-5-(2-methylthioethyl)hydantoin, whereas that from *Arthrobacter aureescens* shows highest activity with arylalkyl substituents, especially 5-benzylhydantoin, at the 5-position [312]. In the enzyme from *Sinorhizobium meliloti*, Cys⁷⁶ is responsible for recognition and proton retrieval of D-isomers, while Cys¹⁸¹ is responsible for L-isomer recognition and racemization [180].
References: [308, 312, 182, 181, 279, 180, 8]

[EC 5.1.99.5 created 2008]

EC 5.2 *cis-trans*-Isomerases

This subclass contains a single sub-subclass for enzymes that rearrange the geometry at double bonds (*cis-trans* isomerases; EC 5.2.1).

EC 5.2.1 *cis-trans* Isomerases (only sub-subclass identified to date)

EC 5.2.1.1

Accepted name: maleate isomerase
Reaction: maleate = fumarate
Systematic name: maleate *cis-trans*-isomerase
References: [25]

[EC 5.2.1.1 created 1961]

EC 5.2.1.2

Accepted name: maleylacetoacetate isomerase
Reaction: 4-maleylacetoacetate = 4-fumarylacetoacetate
Other name(s): maleylacetoacetic isomerase; maleylacetone isomerase; maleylacetone *cis-trans*-isomerase
Systematic name: 4-maleylacetoacetate *cis-trans*-isomerase
Comments: Also acts on maleylpyruvate.
References: [68, 158, 252]

[EC 5.2.1.2 created 1961]

EC 5.2.1.3

Accepted name: retinal isomerase
Reaction: *all-trans*-retinal = 11-*cis*-retinal
Other name(s): retinene isomerase; retinoid isomerase
Systematic name: *all-trans*-retinal 11-*cis-trans*-isomerase
Comments: Light shifts the reaction towards the *cis*-isomer.
References: [119, 255]

[EC 5.2.1.3 created 1961, modified 1976]

EC 5.2.1.4

Accepted name: maleylpyruvate isomerase
Reaction: 3-maleylpyruvate = 3-fumarylpyruvate
Systematic name: 3-maleylpyruvate *cis-trans*-isomerase
References: [158]

[EC 5.2.1.4 created 1965]

EC 5.2.1.5

Accepted name: linoleate isomerase
Reaction: 9-*cis*,12-*cis*-octadecadienoate = 9-*cis*,11-*trans*-octadecadienoate
Other name(s): linoleic acid isomerase
Systematic name: linoleate Δ^{12} -*cis*- Δ^{11} -*trans*-isomerase
References: [141]

[EC 5.2.1.5 created 1972]

EC 5.2.1.6

Accepted name: furylfuramide isomerase
Reaction: (*E*)-2-(2-furyl)-3-(5-nitro-2-furyl)acrylamide = (*Z*)-2-(2-furyl)-3-(5-nitro-2-furyl)acrylamide
Systematic name: 2-(2-furyl)-3-(5-nitro-2-furyl)acrylamide *cis-trans*-isomerase
Comments: Requires NADH.
References: [291]

[EC 5.2.1.6 created 1978]

EC 5.2.1.7

Accepted name: retinol isomerase
Reaction: *all-trans*-retinol = 11-*cis*-retinol
Other name(s): *all-trans*-retinol isomerase
Systematic name: *all-trans*-retinol 11-*cis-trans*-isomerase
Comments: Converts *all-trans*-retinol to 11-*cis*-retinol in the dark, thus reversing the effect of EC 5.2.1.3 retinal isomerase.
References: [29, 38]

[EC 5.2.1.7 created 1989]

EC 5.2.1.8

Accepted name: peptidylprolyl isomerase
Reaction: peptidylproline ($\omega=180$) = peptidylproline ($\omega=0$)
Other name(s): PPIase; cyclophilin [misleading, see comments]; peptide bond isomerase; peptidyl-prolyl *cis-trans* isomerase
Systematic name: peptidylproline *cis-trans*-isomerase
Comments: The first type of this enzyme found [79] proved to be the protein cyclophilin, which binds the immunosuppressant cyclosporin A. Other distinct families of the enzyme exist, one being FK-506 binding proteins (FKBP) and another that includes parvulin from *Escherichia coli*. The three families are structurally unrelated and can be distinguished by being inhibited by cyclosporin A, FK-506 and 5-hydroxy-1,4-naphthoquinone, respectively.
References: [79, 80, 81, 280, 112, 78, 106, 71]

[EC 5.2.1.8 created 1989, modified 2002]

EC 5.2.1.9

Accepted name: farnesol 2-isomerase
Reaction: 2-*trans*,6-*trans*-farnesol = 2-*cis*,6-*trans*-farnesol
Other name(s): farnesol isomerase
Systematic name: 2-*trans*,6-*trans*-farnesol 2-*cis*-*trans*-isomerase
References: [10]

[EC 5.2.1.9 created 1989]

EC 5.2.1.10

Accepted name: 2-chloro-4-carboxymethylenebut-2-en-1,4-olide isomerase
Reaction: *cis*-2-chloro-4-carboxymethylenebut-2-en-1,4-olide = *trans*-2-chloro-4-carboxymethylenebut-2-en-1,4-olide
Other name(s): 2-chlorocarboxymethylenebutenolide isomerase; chlorodienelactone isomerase
Systematic name: 2-chloro-4-carboxymethylenebut-2-en-1,4-olide *cis-trans*-isomerase
References: [249]

[EC 5.2.1.10 created 1992]

[5.2.1.11 Deleted entry. 4-hydroxyphenylacetaldehyde-oxime isomerase. The existence of this enzyme has been called into question by one of the authors of the reference cited]

[EC 5.2.1.11 created 1992, deleted 2005]

EC 5.3 Intramolecular oxidoreductases

These enzymes bring about the oxidation of one part of a molecule with a corresponding reduction of another part. They include the enzymes interconverting, in the sugar series, aldoses and ketoses, and related compounds (sugar isomerases, EC 5.3.1), enzymes catalysing a keto-enol equilibrium (tautomerases, EC 5.3.2), enzymes shifting a carbon-carbon double bond from one position to another (EC 5.3.3), enzymes transposing S-S bonds (EC 5.3.4), and a group of miscellaneous enzymes (EC 5.3.99).

EC 5.3.1 Interconverting aldoses and ketoses, and related compounds

EC 5.3.1.1

Accepted name: triose-phosphate isomerase
Reaction: D-glyceraldehyde 3-phosphate = glycero phosphate
Other name(s): phosphotriose isomerase; triose phosphoisomerase; triose phosphate mutase; D-glyceraldehyde-3-phosphate ketol-isomerase
Systematic name: D-glyceraldehyde-3-phosphate aldose-ketose-isomerase
References: [193, 194]

[EC 5.3.1.1 created 1961]

[5.3.1.2 Deleted entry. erythrose isomerase]

[EC 5.3.1.2 created 1961, deleted 1976]

EC 5.3.1.3

Accepted name: arabinose isomerase
Reaction: D-arabinose = D-ribose

Other name(s): D-arabinose(L-fucose) isomerase; D-arabinose isomerase; L-fucose isomerase; D-arabinose ketol-isomerase
Systematic name: D-arabinose aldose-ketose-isomerase
Comments: Also acts on L-fucose and, more slowly, on L-galactose and D-altrose.
References: [49, 97]

[EC 5.3.1.3 created 1961]

EC 5.3.1.4

Accepted name: L-arabinose isomerase
Reaction: L-arabinose = L-ribulose
Other name(s): L-arabinose ketol-isomerase
Systematic name: L-arabinose aldose-ketose-isomerase
References: [109, 205]

[EC 5.3.1.4 created 1961]

EC 5.3.1.5

Accepted name: xylose isomerase
Reaction: D-xylose = D-xylulose
Other name(s): D-xylose isomerase; D-xylose ketoisomerase; D-xylose ketol-isomerase
Systematic name: D-xylose aldose-ketose-isomerase
Comments: Some enzymes also convert D-glucose to D-fructose.
References: [114, 263, 330]

[EC 5.3.1.5 created 1961 (EC 5.3.1.18 created 1972, part incorporated 1978)]

EC 5.3.1.6

Accepted name: ribose-5-phosphate isomerase
Reaction: D-ribose 5-phosphate = D-ribulose 5-phosphate
Other name(s): phosphopentosisomerase; phosphoriboisomerase; ribose phosphate isomerase; 5-phosphoribose isomerase; D-ribose 5-phosphate isomerase; D-ribose-5-phosphate ketol-isomerase
Systematic name: D-ribose-5-phosphate aldose-ketose-isomerase
Comments: Also acts on D-ribose 5-diphosphate and D-ribose 5-triphosphate.
References: [64, 115, 122]

[EC 5.3.1.6 created 1961]

EC 5.3.1.7

Accepted name: mannose isomerase
Reaction: D-mannose = D-fructose
Other name(s): D-mannose isomerase; D-mannose ketol-isomerase
Systematic name: D-mannose aldose-ketose-isomerase
Comments: Also acts on D-lyxose and D-rhamnose.
References: [219]

[EC 5.3.1.7 created 1961]

EC 5.3.1.8

Accepted name: mannose-6-phosphate isomerase
Reaction: D-mannose 6-phosphate = D-fructose 6-phosphate
Other name(s): phosphomannose isomerase; phosphohexomutase; phosphohexoisomerase; mannose phosphate isomerase; phosphomannoisomerase; D-mannose-6-phosphate ketol-isomerase

Systematic name: D-mannose-6-phosphate aldose-ketose-isomerase
Comments: A zinc protein.
References: [41, 96, 262]

[EC 5.3.1.8 created 1961, modified 1976]

EC 5.3.1.9

Accepted name: glucose-6-phosphate isomerase
Reaction: D-glucose 6-phosphate = D-fructose 6-phosphate
Other name(s): phosphohexose isomerase; phosphohexomutase; oxoisomerase; hexosephosphate isomerase; phosphosaccharomutase; phosphoglucoisomerase; phosphohexoisomerase; phosphoglucose isomerase; glucose phosphate isomerase; hexose phosphate isomerase; D-glucose-6-phosphate ketol-isomerase
Systematic name: D-glucose-6-phosphate aldose-ketose-isomerase
Comments: Also catalyses the anomerization of D-glucose 6-phosphate.
References: [18, 204, 209, 210, 231, 295]

[EC 5.3.1.9 created 1961, modified 1976 (EC 5.3.1.18 created part 1972, incorporated 1978)]

[5.3.1.10 *Transferred entry. glucosamine-6-phosphate isomerase. Now EC 3.5.99.6, glucosamine-6-phosphate deaminase*]

[EC 5.3.1.10 created 1961, deleted 2000]

[5.3.1.11 *Deleted entry. acetylglucosaminophosphate isomerase*]

[EC 5.3.1.11 created 1961, deleted 1978]

EC 5.3.1.12

Accepted name: glucuronate isomerase
Reaction: D-glucuronate = D-fructuronate
Other name(s): uronic isomerase; uronate isomerase; D-glucuronate isomerase; uronic acid isomerase; D-glucuronate ketol-isomerase
Systematic name: D-glucuronate aldose-ketose-isomerase
Comments: Also converts D-galacturonate to D-tagaturonate.
References: [17, 143]

[EC 5.3.1.12 created 1961]

EC 5.3.1.13

Accepted name: arabinose-5-phosphate isomerase
Reaction: D-arabinose 5-phosphate = D-ribulose 5-phosphate
Other name(s): arabinose phosphate isomerase; phosphoarabinoisomerase; D-arabinose-5-phosphate ketol-isomerase
Systematic name: D-arabinose-5-phosphate aldose-ketose-isomerase
References: [305]

[EC 5.3.1.13 created 1965]

EC 5.3.1.14

Accepted name: L-rhamnose isomerase
Reaction: L-rhamnose = L-rhamnulose
Other name(s): rhamnose isomerase; L-rhamnose ketol-isomerase
Systematic name: L-rhamnose aldose-ketose-isomerase
References: [66]

[EC 5.3.1.14 created 1965]

EC 5.3.1.15

Accepted name: D-lyxose ketol-isomerase
Reaction: D-lyxose = D-xylulose
Other name(s): D-lyxose isomerase; D-lyxose ketol-isomerase
Systematic name: D-lyxose aldose-ketose-isomerase
References: [11]

[EC 5.3.1.15 created 1972]

EC 5.3.1.16

Accepted name: 1-(5-phosphoribosyl)-5-[(5-phosphoribosylamino)methylideneamino]imidazole-4-carboxamide isomerase
Reaction: 1-(5-phosphoribosyl)-5-[(5-phosphoribosylamino)methylideneamino]imidazole-4-carboxamide = 5-[(5-phospho-1-deoxyribulos-1-ylamino)methylideneamino]-1-(5-phosphoribosyl)imidazole-4-carboxamide
Other name(s): *N*-(5'-phospho-D-ribosylformimino)-5-amino-1-(5''-phosphoribosyl)-4-imidazolecarboxamide isomerase; phosphoribosylformiminoaminophosphoribosylimidazolecarboxamide isomerase; *N*-(phosphoribosylformimino) aminophosphoribosylimidazolecarboxamide isomerase; 1-(5-phosphoribosyl)-5-[(5-phosphoribosylamino)methylideneamino]imidazole-4-carboxamide ketol-isomerase
Systematic name: 1-(5-phosphoribosyl)-5-[(5-phosphoribosylamino)methylideneamino]imidazole-4-carboxamide aldose-ketose-isomerase
References: [175]

[EC 5.3.1.16 created 1972, modified 2000]

EC 5.3.1.17

Accepted name: 4-deoxy-L-*threo*-5-hexosulose-uronate ketol-isomerase
Reaction: 4-deoxy-L-*threo*-5-hexosulose uronate = 3-deoxy-D-*glycero*-2,5-hexodiulose-uronate
Other name(s): 4-deoxy-L-*threo*-5-hexulose uronate isomerase
Systematic name: 4-deoxy-L-*threo*-5-hexosulose-uronate aldose-ketose-isomerase
References: [228]

[EC 5.3.1.17 created 1972]

[5.3.1.18 Deleted entry. *glucose isomerase*. Reaction is due to EC 5.3.1.9 *glucose-6-phosphate isomerase*, in the presence of arsenate, or EC 5.3.1.5 *xylose isomerase*]

[EC 5.3.1.18 created 1972, deleted 1978]

[5.3.1.19 Transferred entry. *glucosaminophosphate isomerase*. Now EC 2.6.1.16, *glutamine—fructose-6-phosphate transaminase (isomerizing)*]

[EC 5.3.1.19 created 1972, deleted 1984]

EC 5.3.1.20

Accepted name: ribose isomerase
Reaction: D-ribose = D-ribulose
Other name(s): D-ribose isomerase; D-ribose ketol-isomerase
Systematic name: D-ribose aldose-ketose-isomerase
Comments: Also acts on L-lyxose and L-rhamnose.
References: [124]

[EC 5.3.1.20 created 1978]

EC 5.3.1.21

Accepted name: corticosteroid side-chain-isomerase
Reaction: 11-deoxycorticosterone = 20-hydroxy-3-oxopregn-4-en-21-al
Systematic name: 11-deoxycorticosterone aldose-ketose-isomerase
Comments: An epimerization at C-20 and C-21 is probably catalysed by the same enzyme.
References: [178, 197]

[EC 5.3.1.21 created 1983]

EC 5.3.1.22

Accepted name: hydroxypyruvate isomerase
Reaction: hydroxypyruvate = 2-hydroxy-3-oxopropanoate
Systematic name: hydroxypyruvate aldose-ketose-isomerase
References: [316]

[EC 5.3.1.22 created 1983]

EC 5.3.1.23

Accepted name: *S*-methyl-5-thioribose-1-phosphate isomerase
Reaction: *S*-methyl-5-thio- α -D-ribose 1-phosphate = *S*-methyl-5-thio-D-ribulose 1-phosphate
Other name(s): methylthioribose 1-phosphate isomerase; 1-PMTR isomerase; 5-methylthio-5-deoxy-D-ribose-1-phosphate ketol-isomerase; *S*-methyl-5-thio-5-deoxy-D-ribose-1-phosphate ketol-isomerase; *S*-methyl-5-thio-5-deoxy-D-ribose-1-phosphate aldose-ketose-isomerase; 1-phospho-5'-*S*-methylthioribose isomerase; *S*-methyl-5-thio-D-ribose-1-phosphate aldose-ketose-isomerase
Systematic name: *S*-methyl-5-thio- α -D-ribose-1-phosphate aldose-ketose-isomerase
References: [90, 293, 86]

[EC 5.3.1.23 created 1989]

EC 5.3.1.24

Accepted name: phosphoribosylanthranilate isomerase
Reaction: *N*-(5-phospho- β -D-ribosyl)anthranilate = 1-(2-carboxyphenylamino)-1-deoxy-D-ribulose 5-phosphate
Other name(s): PRA isomerase; PRAI; IGPS:PRAI (indole-3-glycerol-phosphate synthetase/*N*-5'-phosphoribosylanthranilate isomerase complex); *N*-(5-phospho- β -D-ribosyl)anthranilate ketol-isomerase
Systematic name: *N*-(5-phospho- β -D-ribosyl)anthranilate aldose-ketose-isomerase
Comments: In some organisms, this enzyme is part of a multifunctional protein, together with one or more other components of the system for the biosynthesis of tryptophan [EC 2.4.2.18 (anthranilate phosphoribosyltransferase), EC 4.1.1.48 (indole-3-glycerol-phosphate synthase), EC 4.1.3.27 (anthranilate synthase) and EC 4.2.1.20 (tryptophan synthase)].
References: [36, 51, 123]

[EC 5.3.1.24 created 1990]

EC 5.3.1.25

Accepted name: L-fucose isomerase
Reaction: L-fucose = L-fuculose
Systematic name: L-fucose aldose-ketose-isomerase
References: [171]

[EC 5.3.1.25 created 1999]

EC 5.3.1.26

Accepted name: galactose-6-phosphate isomerase
Reaction: D-galactose 6-phosphate = D-tagatose 6-phosphate
Systematic name: D-galactose-6-phosphate aldose-ketose-isomerase
Comments: Involved in the tagatose 6-phosphate pathway of lactose catabolism in bacteria.
References: [306, 241]

[EC 5.3.1.26 created 1999]

EC 5.3.1.27

Accepted name: 6-phospho-3-hexuloisomerase
Reaction: D-*arabino*-hex-3-ulose 6-phosphate = D-fructose 6-phosphate
Other name(s): 3-hexulose-6-phosphate isomerase; phospho-3-hexuloisomerase; PHI; 6-phospho-3-hexulose isomerase; YckF
Systematic name: D-*arabino*-hex-3-ulose-6-phosphate isomerase
Comments: This enzyme, along with EC 4.1.2.43, 3-hexulose-6-phosphate synthase, plays a key role in the ribulose-monophosphate cycle of formaldehyde fixation, which is present in many microorganisms that are capable of utilizing C1-compounds [75]. The hyperthermophilic and anaerobic archaeon *Pyrococcus horikoshii* OT3 constitutively produces a bifunctional enzyme that sequentially catalyses the reactions of EC 4.1.2.43 (3-hexulose-6-phosphate synthase) and this enzyme [213].
References: [75, 339, 136, 213, 179, 283]

[EC 5.3.1.27 created 2008]

EC 5.3.1.28

Accepted name: D-sedoheptulose 7-phosphate isomerase
Reaction: D-sedoheptulose 7-phosphate = D-*glycero*-D-*manno*-heptose 7-phosphate
Other name(s): sedoheptulose-7-phosphate isomerase; phosphoheptose isomerase; *gmhA* (gene name); *lpcA* (gene name)
Systematic name: D-*glycero*-D-*manno*-heptose 7-phosphate aldose-ketose-isomerase
Comments: In Gram-negative bacteria the enzyme is involved in biosynthesis of ADP-L-*glycero*-β-D-*manno*-heptose, which is utilized for assembly of the lipopolysaccharide inner core. In Gram-positive bacteria the enzyme is involved in biosynthesis of GDP-D-*glycero*-α-D-*manno*-heptose, which is required for assembly of S-layer glycoprotein.
References: [150, 149, 301, 144, 285]

[EC 5.3.1.28 created 2010]

EC 5.3.2 Interconverting keto- and enol-groups

EC 5.3.2.1

Accepted name: phenylpyruvate tautomerase
Reaction: *keto*-phenylpyruvate = *enol*-phenylpyruvate
Other name(s): phenylpyruvic keto-enol isomerase
Systematic name: phenylpyruvate *keto*—*enol*-isomerase
Comments: Also acts on other arylpyruvates.
References: [31, 151, 152]

[EC 5.3.2.1 created 1961]

EC 5.3.2.2

Accepted name: oxaloacetate tautomerase

Reaction: *keto*-oxaloacetate = *enol*-oxaloacetate
Other name(s): oxalacetic keto-enol isomerase
Systematic name: oxaloacetate *keto*—*enol*-isomerase
References: [13]

[EC 5.3.2.2 created 1972]

EC 5.3.3 Transposing C=C bonds

EC 5.3.3.1

Accepted name: steroid Δ -isomerase
Reaction: a 3-oxo- Δ^5 -steroid = a 3-oxo- Δ^4 -steroid
Other name(s): hydroxysteroid isomerase; steroid isomerase; Δ^5 -ketosteroid isomerase; Δ^5 (or Δ^4)-3-keto steroid isomerase; Δ^5 -steroid isomerase; 3-oxosteroid isomerase; Δ^5 -3-keto steroid isomerase; Δ^5 -3-oxosteroid isomerase
Systematic name: 3-oxosteroid Δ^5 - Δ^4 -isomerase
Comments: May be at least three distinct enzymes.
References: [72, 137, 281]

[EC 5.3.3.1 created 1961]

EC 5.3.3.2

Accepted name: isopentenyl-diphosphate Δ -isomerase
Reaction: isopentenyl diphosphate = dimethylallyl diphosphate
Other name(s): isopentenylpyrophosphate Δ -isomerase; methylbutenylpyrophosphate isomerase; isopentenylpyrophosphate isomerase
Systematic name: isopentenyl-diphosphate Δ^3 - Δ^2 -isomerase
Comments: The enzyme from *Streptomyces* sp. strain CL190 requires FMN and NAD(P)H as cofactors. Activity is reduced if FMN is replaced by FAD, but the enzyme becomes inactive when NAD(P)H is replaced by NAD⁺ or NADP⁺. That enzyme also requires Mg²⁺, Mn²⁺ or Ca²⁺ for activity.
References: [134, 30, 3]

[EC 5.3.3.2 created 1961, modified 2002]

EC 5.3.3.3

Accepted name: vinylacetyl-CoA Δ -isomerase
Reaction: vinylacetyl-CoA = crotonyl-CoA
Other name(s): vinylacetyl coenzyme A Δ -isomerase; vinylacetyl coenzyme A isomerase; Δ^3 -*cis*- Δ^2 -*trans*-enoyl-CoA isomerase
Systematic name: vinylacetyl-CoA Δ^3 - Δ^2 -isomerase
Comments: Also acts on 3-methyl-vinylacetyl-CoA.
References: [172, 239]

[EC 5.3.3.3 created 1961]

EC 5.3.3.4

Accepted name: muconolactone Δ -isomerase
Reaction: (*S*)-5-oxo-2,5-dihydrofuran-2-acetate = 5-oxo-4,5-dihydrofuran-2-acetate
Other name(s): muconolactone isomerase
Systematic name: 5-oxo-4,5-dihydrofuran-2-acetate Δ^3 - Δ^2 -isomerase
References: [214, 216]

[EC 5.3.3.4 created 1961 as EC 3.1.1.16, part transferred 1972 to EC 5.3.3.4 rest to EC 5.3.3.4]

EC 5.3.3.5

Accepted name: cholestenol Δ -isomerase
Reaction: 5α -cholest-7-en-3 β -ol = 5α -cholest-8-en-3 β -ol
Systematic name: Δ^7 -cholestenol Δ^7 - Δ^8 -isomerase
References: [315]

[EC 5.3.3.5 created 1972]

EC 5.3.3.6

Accepted name: methylitaconate Δ -isomerase
Reaction: methylitaconate = 2,3-dimethylmaleate
Other name(s): methylitaconate isomerase
Systematic name: methylitaconate Δ^2 - Δ^3 -isomerase
References: [156]

[EC 5.3.3.6 created 1972]

EC 5.3.3.7

Accepted name: aconitate Δ -isomerase
Reaction: *trans*-aconitate = *cis*-aconitate
Other name(s): aconitate isomerase
Systematic name: aconitate Δ^2 - Δ^3 -isomerase
Comments: *cis*-Aconitate is used to designate the isomer (*Z*)-prop-1-ene-1,2,3-tricarboxylate. This isomerization could take place either in a direct *cis-trans* interconversion or by an allelic rearrangement; the enzyme has been shown to catalyse the latter change.
References: [148, 147]

[EC 5.3.3.7 created 1972]

EC 5.3.3.8

Accepted name: dodecenoyl-CoA isomerase
Reaction: (3*Z*)-dodec-3-enoyl-CoA = (2*E*)-dodec-2-enoyl-CoA
Other name(s): dodecenoyl-CoA Δ -isomerase; Δ^3 -*cis*- Δ^2 -*trans*-enoyl-CoA isomerase; acetylene-allene isomerase; dodecenoyl-CoA Δ -isomerase; dodecenoyl-CoA Δ^3 -*cis*- Δ^2 -*trans*-isomerase
Systematic name: dodecenoyl-CoA (3*Z*)-(2*E*)-isomerase
Comments: Also catalyses the interconversion of 3-acetylenic fatty acyl thioesters and (+)-2,3-dienoyl fatty acyl thioesters, with fatty acid chain lengths C₆ to C₁₂.
References: [195, 273, 274, 275]

[EC 5.3.3.8 created 1978, modified 1980]

EC 5.3.3.9

Accepted name: prostaglandin-A₁ Δ -isomerase
Reaction: (13*E*)-(15*S*)-15-hydroxy-9-oxoprostano-10,13-dienoate = (13*E*)-(15*S*)-15-hydroxy-9-oxoprostano-11,13-dienoate
Other name(s): prostaglandin A isomerase
Systematic name: (13*E*)-(15*S*)-15-hydroxy-9-oxoprostano-10,13-dienoate Δ^{10} - Δ^{11} -isomerase
Comments: Interconverts prostaglandin A₁ and prostaglandin C₁.
References: [102]

[EC 5.3.3.9 created 1978]

EC 5.3.3.10

Accepted name: 5-carboxymethyl-2-hydroxyumuconate Δ -isomerase
Reaction: 5-carboxymethyl-2-hydroxyumuconate = 5-carboxy-2-oxohept-3-enedioate
Systematic name: 5-carboxymethyl-2-hydroxyumuconate Δ^2, Δ^4 -2-oxo, Δ^3 -isomerase
References: [87]

[EC 5.3.3.10 created 1984]

EC 5.3.3.11

Accepted name: isopiperitenone Δ -isomerase
Reaction: isopiperitenone = piperitenone
Systematic name: isopiperitenone Δ^8 - Δ^4 -isomerase
Comments: Involved in the biosynthesis of menthol and related monoterpenes in peppermint (*Mentha piperita*) leaves.
References: [146]

[EC 5.3.3.11 created 1989]

EC 5.3.3.12

Accepted name: L-dopachrome isomerase
Reaction: L-dopachrome = 5,6-dihydroxyindole-2-carboxylate
Other name(s): dopachrome tautomerase; tyrosinase-related protein 2; TRP-1; TRP2; TRP-2; tyrosinase-related protein-2; dopachrome Δ^7, Δ^2 -isomerase; dopachrome Δ -isomerase; dopachrome conversion factor; dopachrome isomerase; dopachrome oxidoreductase; dopachrome-rearranging enzyme; DCF; DCT; dopachrome keto-enol isomerase; L-dopachrome-methyl ester tautomerase
Systematic name: L-dopachrome keto-enol isomerase
Comments: A zinc enzyme. Stereospecific for L-dopachrome. Dopachrome methyl ester is a substrate, but dopaminochrome (2,3-dihydroindole-5,6-quinone) is not (see also EC 4.1.1.84, D-dopachrome decarboxylase).
References: [266, 220, 221]

[EC 5.3.3.12 created 1992, modified 1999, modified 2005]

EC 5.3.3.13

Accepted name: polyenoic fatty acid isomerase
Reaction: (5Z,8Z,11Z,14Z,17Z)-icosapentaenoate = (5Z,7E,9E,14Z,17Z)-icosapentaenoate
Other name(s): PFI; eicosapentaenoate *cis*- $\Delta^{5,8,11,14,17}$ -eicosapentaenoate *cis*- Δ^5 -*trans*- $\Delta^{7,9}$ -*cis*- $\Delta^{14,17}$ isomerase; (5Z,8Z,11Z,14Z,17Z)-eicosapentaenoate $\Delta^{8,11}$ - $\Delta^{7,8}$ -isomerase (incorrect); (5Z,8Z,11Z,14Z,17Z)-eicosapentaenoate $\Delta^{8,11}$ - $\Delta^{7,9}$ -isomerase (*trans*-double-bond-forming)
Systematic name: (5Z,8Z,11Z,14Z,17Z)-icosapentaenoate $\Delta^{8,11}$ - $\Delta^{7,9}$ -isomerase (*trans*-double-bond-forming)
Comments: The enzyme from the red alga *Ptilota filicina* catalyses the isomerization of skip dienes (methylene-interrupted double bonds) in a broad range of fatty acids and fatty-acid analogues, such as arachidonate and γ -linolenate, to yield a conjugated triene.
References: [317, 319, 318, 341]

[EC 5.3.3.13 created 2004]

EC 5.3.3.14

Accepted name: *trans*-2-decenoyl-[acyl-carrier protein] isomerase
Reaction: a *trans*-dec-2-enoyl-[acyl-carrier protein] = a *cis*-dec-3-enoyl-[acyl-carrier protein]

Other name(s): β -hydroxydecanoyl thioester dehydrase; *trans*-2-*cis*-3-decenoyl-ACP isomerase; *trans*-2,*cis*-3-decenoyl-ACP isomerase; *trans*-2-decenoyl-ACP isomerase; FabM; decenoyl-[acyl-carrier-protein] Δ^2 -*trans*- Δ^3 -*cis*-isomerase

Systematic name: decenoyl-[acyl-carrier protein] Δ^2 -*trans*- Δ^3 -*cis*-isomerase

Comments: While the enzyme from *Escherichia coli* is highly specific for the 10-carbon enoyl-ACP, the enzyme from *Streptococcus pneumoniae* can also use the 12-carbon enoyl-ACP as substrate in vitro but not 14- or 16-carbon enoyl-ACPs [177]. ACP can be replaced by either CoA or *N*-acetylcysteamine thioesters. The *cis*-3-enoyl product is required to form unsaturated fatty acids, such as palmitoleic acid and *cis*-vaccenic acid, in dissociated (or type II) fatty-acid biosynthesis.

References: [39, 32, 177, 52]

[EC 5.3.3.14 created 2006]

EC 5.3.3.15

Accepted name: ascopyrone tautomerase

Reaction: 1,5-anhydro-4-deoxy-D-*glycero*-hex-3-en-2-ulose = 1,5-anhydro-4-deoxy-D-*glycero*-hex-1-en-3-ulose

Other name(s): ascopyrone isomerase; ascopyrone intramolecular oxidoreductase; 1,5-anhydro-D-*glycero*-hex-3-en-2-ulose tautomerase; APM tautomerase; ascopyrone P tautomerase; APTM

Systematic name: 1,5-anhydro-4-deoxy-D-*glycero*-hex-3-en-2-ulose Δ^3 - Δ^1 -isomerase

Comments: This enzyme catalyses one of the steps in the anhydrofructose pathway, which leads to the degradation of glycogen and starch via 1,5-anhydro-D-fructose [338, 337]. The other enzymes involved in this pathway are EC 4.2.1.110 (aldos-2-ulose dehydratase), EC 4.2.1.111 (1,5-anhydro-D-fructose dehydratase) and EC 4.2.2.13 [exo-(1 \rightarrow 4)- α -D-glucan lyase]. Ascopyrone P is an anti-oxidant [337].

References: [338, 337]

[EC 5.3.3.15 created 2006]

EC 5.3.4 Transposing S-S bonds

EC 5.3.4.1

Accepted name: protein disulfide-isomerase

Reaction: Catalyses the rearrangement of -S-S- bonds in proteins

Other name(s): S-S rearrangase

Systematic name: protein disulfide-isomerase

Comments: Needs reducing agents or partly reduced enzyme; the reaction depends on sulfhydryl-disulfide interchange.

References: [170, 84]

[EC 5.3.4.1 created 1972]

EC 5.3.99 Other intramolecular oxidoreductases

[5.3.99.1 Deleted entry. *hydroperoxide isomerase*. Reaction due to combined action of EC 4.2.1.92 (*hydroperoxide dehydratase*) and EC 5.3.99.6 (*allene-oxide cyclase*)]

[EC 5.3.99.1 created 1972, deleted 1992]

EC 5.3.99.2

Accepted name: prostaglandin-D synthase

Reaction: (5*Z*,13*E*,15*S*)-9 α ,11 α -epidioxy-15-hydroxyprosta-5,13-dienoate = (5*Z*,13*E*,15*S*)-9 α ,15-dihydroxy-11-oxoprosta-5,13-dienoate

Other name(s): prostaglandin-H₂ Δ-isomerase; prostaglandin-R-prostaglandin D isomerase; PGH-PGD isomerase(5,13)-(15S)-9α,11α-epidioxy-15-hydroxyprosta-5,13-dienoate Δ-isomerase (incorrect); (5,13)-(15S)-9α,11α-epidioxy-15-hydroxyprosta-5,13-dienoate D-isomerase; prostaglandin endoperoxide Δ-isomerase; prostaglandin D synthetase
Systematic name: (5Z,13E,15S)-9α,11α-epidioxy-15-hydroxyprosta-5,13-dienoate D-isomerase
Comments: Brings about the opening of the epidioxy bridge. Some enzymes require glutathione.
References: [48, 257]

[EC 5.3.99.2 created 1976, modified 1990]

EC 5.3.99.3

Accepted name: prostaglandin-E synthase
Reaction: (5Z,13E)-(15S)-9α,11α-epidioxy-15-hydroxyprosta-5,13-dienoate = (5Z,13E)-(15S)-11α,15-dihydroxy-9-oxoprosta-5,13-dienoate
Other name(s): prostaglandin-H₂ E-isomerase; endoperoxide isomerase; endoperoxide isomerase; prostaglandin R-prostaglandin E isomerase; prostaglandin endoperoxide E isomerase; PGE isomerase; PGH-PGE isomerase; PGE₂ isomerase; prostaglandin endoperoxide E₂ isomerase; prostaglandin H-E isomerase
Systematic name: (5Z,13E)-(15S)-9α,11α-epidioxy-15-hydroxyprosta-5,13-dienoate E-isomerase
Comments: Brings about the opening of the epidioxy bridge. Requires glutathione.
References: [211, 282]

[EC 5.3.99.3 created 1976, modified 1990]

EC 5.3.99.4

Accepted name: prostaglandin-I synthase
Reaction: (5Z,13E)-(15S)-9α,11α-epidioxy-15-hydroxyprosta-5,13-dienoate = (5Z,13E)-(15S)-6,9α-epoxy-11α,15-dihydroxyprosta-5,13-dienoate
Other name(s): prostacyclin synthase; prostacycline synthetase; prostagladin I₂ synthetase; PGI₂ synthase; PGI₂ synthetase
Systematic name: (5Z,13E)-(15S)-9α,11α-epidioxy-15-hydroxyprosta-5,13-dienoate 6-isomerase
Comments: Converts prostaglandin H₂ into prostaglandin I₂ (prostacyclin). A heme-thiolate protein.
References: [63, 297]

[EC 5.3.99.4 created 1984, modified 1990]

EC 5.3.99.5

Accepted name: thromboxane-A synthase
Reaction: (5Z,13E)-(15S)-9α,11α-epidioxy-15-hydroxyprosta-5,13-dienoate = (5Z,13E)-(15S)-9α,11α-epoxy-15-hydroxythromboxa-5,13-dienoate
Other name(s): thromboxane synthase; (5Z,13E)-(15S)-9α,11α-epidioxy-15-hydroxyprosta-5,13-dienoate thromboxane-A₂-isomerase
Systematic name: (5Z,13E)-(15S)-9α,11α-epidioxy-15-hydroxyprosta-5,13-dienoate isomerase
Comments: Converts prostaglandin H₂ into thromboxane A₂. A heme-thiolate protein.
References: [253, 298]

[EC 5.3.99.5 created 1984, modified 1990]

EC 5.3.99.6

Accepted name: allene-oxide cyclase
Reaction: (9Z)-(13S)-12,13-epoxyoctadeca-9,11,15-trienoate = (15Z)-12-oxophyto-10,15-dienoate
Systematic name: (9Z)-(13S)-12,13-epoxyoctadeca-9,11,15-trienoate isomerase (cyclizing)
Comments: Allene oxides formed by the action of EC 4.2.1.92 hydroperoxide dehydratase, are converted into cyclopentenone derivatives.
References: [104]

[EC 5.3.99.6 created 1992]

EC 5.3.99.7

Accepted name: styrene-oxide isomerase
Reaction: styrene oxide = phenylacetaldehyde
Other name(s): SOI
Systematic name: styrene-oxide isomerase (epoxide-cleaving)
Comments: Highly specific.
References: [107]

[EC 5.3.99.7 created 1992]

EC 5.3.99.8

Accepted name: capsanthin/capsorubin synthase
Reaction: (1) violaxanthin = capsorubin
(2) antheraxanthin = capsanthin
Other name(s): CCS; ketoxanthophyll synthase; capsanthin-capsorubin synthase
Systematic name: violaxanthin—capsorubin isomerase (ketone-forming)
Comments: This multifunctional enzyme is induced during chromoplast differentiation in plants [34]. Isomerization of the epoxide ring of violaxanthin gives the cyclopentyl-ketone of capsorubin or capsanthin.
References: [34, 163, 327]

[EC 5.3.99.8 created 2005]

EC 5.3.99.9

Accepted name: neoxanthin synthase
Reaction: violaxanthin = neoxanthin
Other name(s): NSY
Systematic name: violaxanthin—neoxanthin isomerase (epoxide-opening)
Comments: The opening of the epoxide ring of violaxanthin generates a chiral allene. Neoxanthin is a precursor of the plant hormone abscisic acid and the last product of carotenoid synthesis in green plants [33].
References: [6, 33]

[EC 5.3.99.9 created 2005]

EC 5.4 Intramolecular transferases

This subclass contains enzymes that transfer a group from one position to another within a molecule. Sub-subclasses are based on the group transferred: acyl group (EC 5.4.1), phospho group (EC 5.4.2), amino group (EC 5.4.3), hydroxy group (EC 5.4.4), or some other group (EC 5.4.99).

EC 5.4.1 Transferring acyl groups

EC 5.4.1.1

Accepted name: lysolecithin acylmutase
Reaction: 2-lysolecithin = 3-lysolecithin
Other name(s): lysolecithin migratase
Systematic name: lysolecithin 2,3-acylmutase
References: [300]

[EC 5.4.1.1 created 1961]

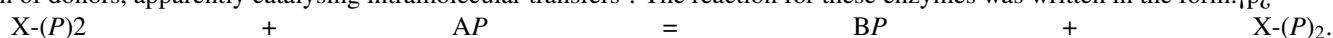
EC 5.4.1.2

Accepted name: precorrin-8X methylmutase
Reaction: precorrin-8X = hydrogenobyryinate
Other name(s): precorrin isomerase; hydrogenobyrynic acid-binding protein
Systematic name: precorrin-8X 11,12-methylmutase
References: [287, 244, 240, 54]

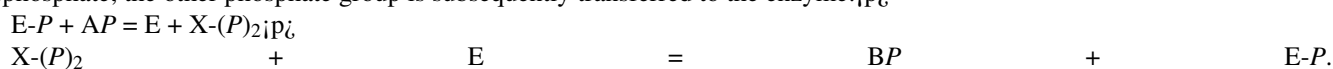
[EC 5.4.1.2 created 1999]

EC 5.4.2 Phosphotransferases (phosphomutases)

Most of these enzymes were previously listed as sub-subclass EC 2.7.5, under the heading: 'Phosphotransferases with regeneration of donors, apparently catalysing intramolecular transfers'. The reaction for these enzymes was written in the form:



In fact, since phosphorylation of the acceptor produces a bisphosphate that is identical to the donor, the overall reaction is an isomerization of AP into BP, with the bisphosphate acting catalytically. It has been shown in some cases that the enzyme has a functional phosphate group, which can act as the donor. Phosphate is transferred to the substrate, forming the intermediate bisphosphate; the other phosphate group is subsequently transferred to the enzyme:



The bisphosphate may be firmly attached to the enzyme during the catalytic cycle, or, in other cases, may be released so that free bisphosphate is required as an activator. Under these circumstances, it was agreed in 1983 that all of these enzymes should be listed together in this sub-subclass based on the overall isomerase reaction.

EC 5.4.2.1

Accepted name: phosphoglycerate mutase
Reaction: 2-phospho-D-glycerate = 3-phospho-D-glycerate
Other name(s): phosphoglycerate phosphomutase; phosphoglyceromutase; glycerate phosphomutase (diphosphoglycerate cofactor); monophosphoglycerate mutase; monophosphoglyceromutase; diphosphoglycomutase; diphosphoglycerate mutase; bisphosphoglyceromutase; GriP mutase; MPGM; PGA mutase; PGAM-i; PGAM; PGAM-d; PGM
Systematic name: D-phosphoglycerate 2,3-phosphomutase
Comments: The enzymes from mammals and from yeast are phosphorylated by (2R)-2,3-bis-phosphoglycerate, which is also an intermediate (see introduction to section EC 5.4.2). With the rabbit muscle enzyme, dissociation of bisphosphate from the enzyme is much slower than the overall isomerization. These enzymes also catalyse, slowly, the reactions of EC 5.4.2.4 bisphosphoglycerate mutase; they were formerly listed as EC 2.7.5.3. Enzymes from wheat, rice, insects and some fungi, however, have maximum activity in the absence of 2,3-bisphosphoglycerate, and were formerly listed under the present number as phosphoglycerate phosphomutase.
References: [99, 234, 243]

[EC 5.4.2.1 created 1961 (EC 2.7.5.3 created 1961, incorporated 1984)]

EC 5.4.2.2

Accepted name: phosphoglucomutase
Reaction: α -D-glucose 1-phosphate = D-glucose 6-phosphate
Other name(s): glucose phosphomutase; phosphoglucose mutase
Systematic name: α -D-glucose 1,6-phosphomutase

Comments: Maximum activity is only obtained in the presence of α -D-glucose 1,6-bisphosphate. This bisphosphate is an intermediate in the reaction, being formed by transfer of a phosphate residue from the enzyme to the substrate, but the dissociation of bisphosphate from the enzyme complex is much slower than the overall isomerization. The enzyme also catalyses (more slowly) the interconversion of 1-phosphate and 6-phosphate isomers of many other α -D-hexoses, and the interconversion of α -D-ribose 1-phosphate and 5-phosphate.

References: [130, 201, 235, 234, 278]

[EC 5.4.2.2 created 1961 as EC 2.7.5.1, transferred 1984 to EC 5.4.2.2]

EC 5.4.2.3

Accepted name: phosphoacetylglucosamine mutase

Reaction: *N*-acetyl- α -D-glucosamine 1-phosphate = *N*-acetyl-D-glucosamine 6-phosphate

Other name(s): acetylglucosamine phosphomutase; acetylglucosamine phosphomutase; acetylaminodeoxyglucose phosphomutase; phospho-*N*-acetylglucosamine mutase; *N*-acetyl-D-glucosamine 1,6-phosphomutase

Systematic name: *N*-acetyl- α -D-glucosamine 1,6-phosphomutase

Comments: The enzyme is activated by *N*-acetyl- α -D-glucosamine 1,6-bisphosphate.

References: [44, 165, 234, 237]

[EC 5.4.2.3 created 1961 as EC 2.7.5.2, transferred 1984 to EC 5.4.2.3]

EC 5.4.2.4

Accepted name: bisphosphoglycerate mutase

Reaction: 3-phospho-D-glyceroyl phosphate = 2,3-bisphospho-D-glycerate

Other name(s): diphosphoglycerate mutase; glycerate phosphomutase; bisphosphoglycerate synthase; bisphosphoglyceromutase; biphosphoglycerate synthase; diphosphoglyceric mutase; 2,3-diphosphoglycerate mutase; phosphoglyceromutase; 2,3-diphosphoglycerate synthase; DPGM; 2,3-bisphosphoglycerate mutase; BPGM; diphosphoglyceromutase; 2,3-diphosphoglyceromutase

Systematic name: 3-phospho-D-glycerate 1,2-phosphomutase

Comments: In the direction shown, this enzyme is phosphorylated by 3-phosphoglyceroyl phosphate, to give phosphoenzyme and 3-phosphoglycerate. The latter is rephosphorylated by the enzyme to yield 2,3-bisphosphoglycerate, but this reaction is slowed by dissociation of 3-phosphoglycerate from the enzyme, which is therefore more active in the presence of added 3-phosphoglycerate. This enzyme also catalyses, slowly, the reactions of EC 3.1.3.13 (bisphosphoglycerate phosphatase) and EC 5.4.2.1 (phosphoglycerate mutase).

References: [234, 242, 243]

[EC 5.4.2.4 created 1961 as EC 2.7.5.4, transferred 1984 to EC 5.4.2.4]

EC 5.4.2.5

Accepted name: phosphoglucomutase (glucose-cofactor)

Reaction: α -D-glucose 1-phosphate = D-glucose 6-phosphate

Other name(s): glucose phosphomutase; glucose-1-phosphate phosphotransferase

Systematic name: α -D-glucose 1,6-phosphomutase (glucose-cofactor)

Comments: The enzyme is activated by D-glucose, which probably acts as an acceptor for a phosphate residue from the substrate, thus being itself converted into the product.

References: [85, 234]

[EC 5.4.2.5 created 1972 as EC 2.7.5.5, transferred 1984 to EC 5.4.2.5]

EC 5.4.2.6

Accepted name: β -phosphoglucomutase

Reaction: β -D-glucose 1-phosphate = β -D-glucose 6-phosphate

Systematic name: β -D-glucose 1,6-phosphomutase
Comments: No cofactor requirement has been demonstrated.
References: [26, 234]

[EC 5.4.2.6 created 1984]

EC 5.4.2.7

Accepted name: phosphopentomutase
Reaction: α -D-ribose 1-phosphate = D-ribose 5-phosphate
Other name(s): phosphodeoxyribomutase; deoxyribose phosphomutase; deoxyribomutase; phosphoribomutase; α -D-glucose-1,6-bisphosphate:deoxy-D-ribose-1-phosphate phosphotransferase; D-ribose 1,5-phosphomutase
Systematic name: α -D-ribose 1,5-phosphomutase
Comments: Also converts 2-deoxy- α -D-ribose 1-phosphate into 2-deoxy-D-ribose 5-phosphate. α -D-Ribose 1,5-bisphosphate, 2-deoxy- α -D-ribose 1,5-bisphosphate, or α -D-glucose 1,6-bisphosphate can act as cofactor.
References: [105, 132, 234]

[EC 5.4.2.7 created 1972 as EC 2.7.5.6, transferred 1984 to EC 5.4.2.7]

EC 5.4.2.8

Accepted name: phosphomannomutase
Reaction: α -D-mannose 1-phosphate = D-mannose 6-phosphate
Other name(s): mannose phosphomutase; phosphomannose mutase; D-mannose 1,6-phosphomutase
Systematic name: α -D-mannose 1,6-phosphomutase
Comments: α -D-Mannose 1,6-bisphosphate or α -D-glucose 1,6-bisphosphate can act as cofactor.
References: [264]

[EC 5.4.2.8 created 1981 as EC 2.7.5.7, transferred 1984 to EC 5.4.2.8]

EC 5.4.2.9

Accepted name: phospho*enol*pyruvate mutase
Reaction: phospho*enol*pyruvate = 3-phosphonopyruvate
Other name(s): phospho*enol*pyruvate-phosphonopyruvate phosphomutase; PEP phosphomutase; phospho*enol*pyruvate phosphomutase; PEPPM; PEP phosphomutase
Systematic name: phospho*enol*pyruvate 2,3-phosphonomutase
Comments: Involved in the biosynthesis of the C-P bond, although the equilibrium greatly favours phospho*enol*pyruvate.
References: [35, 113, 251]

[EC 5.4.2.9 created 1990]

EC 5.4.2.10

Accepted name: phosphoglucosamine mutase
Reaction: α -D-glucosamine 1-phosphate = D-glucosamine 6-phosphate
Systematic name: α -D-glucosamine 1,6-phosphomutase
Comments: The enzyme is involved in the pathway for bacterial cell-wall peptidoglycan and lipopolysaccharide biosyntheses, being an essential step in the pathway for UDP-*N*-acetylglucosamine biosynthesis. The enzyme from *Escherichia coli* is activated by phosphorylation and can be autophosphorylated in vitro by α -D-glucosamine 1,6-bisphosphate, which is an intermediate in the reaction, α -D-glucose 1,6-bisphosphate or ATP. It can also catalyse the interconversion of α -D-glucose 1-phosphate and glucose 6-phosphate, although at a much lower rate.
References: [190, 60, 129, 127, 128]

[EC 5.4.2.10 created 2001]

EC 5.4.3 Transferring amino groups

[5.4.3.1 Deleted entry. ornithine 4,5-aminomutase. This reaction was due to a mixture of EC 5.1.1.12 (ornithine racemase) and EC 5.4.3.5 (D-ornithine 4,5-aminomutase)]

[EC 5.4.3.1 created 1972, deleted 1976]

EC 5.4.3.2

Accepted name: lysine 2,3-aminomutase
Reaction: L-lysine = (3*S*)-3,6-diaminohexanoate
Systematic name: L-lysine 2,3-aminomutase
Comments: Activity is stimulated by *S*-adenosyl-L-methionine and pyridoxal phosphate.
References: [1, 340]

[EC 5.4.3.2 created 1972]

EC 5.4.3.3

Accepted name: β-lysine 5,6-aminomutase
Reaction: (3*S*)-3,6-diaminohexanoate = (3*S*,5*S*)-3,5-diaminohexanoate
Other name(s): β-lysine mutase; L-β-lysine 5,6-aminomutase
Systematic name: (3*S*)-3,6-diaminohexanoate 5,6-aminomutase
Comments: Requires a cobamide coenzyme.
References: [238, 270]

[EC 5.4.3.3 created 1972]

EC 5.4.3.4

Accepted name: D-lysine 5,6-aminomutase
Reaction: D-lysine = 2,5-diaminohexanoate
Other name(s): D-α-lysine mutase; adenosylcobalamin-dependent D-lysine 5,6-aminomutase
Systematic name: D-2,6-diaminohexanoate 5,6-aminomutase
Comments: Requires a cobamide coenzyme.
References: [198, 271]

[EC 5.4.3.4 created 1972, modified 2003]

EC 5.4.3.5

Accepted name: D-ornithine 4,5-aminomutase
Reaction: D-ornithine = (2*R*,4*S*)-2,4-diaminopentanoate
Other name(s): D-α-ornithine 5,4-aminomutase; D-ornithine aminomutase
Systematic name: D-ornithine 4,5-aminomutase
Comments: A pyridoxal-phosphate protein that requires a cobamide coenzyme for activity.
References: [267]

[EC 5.4.3.5 created 1972 as EC 5.4.3.1, transferred 1976 to EC 5.4.3.5, modified 2003]

EC 5.4.3.6

Accepted name: tyrosine 2,3-aminomutase
Reaction: L-tyrosine = 3-amino-3-(4-hydroxyphenyl)propanoate
Other name(s): tyrosine α,β-mutase

Systematic name: L-tyrosine 2,3-aminomutase
Comments: Requires ATP.
References: [157]

[EC 5.4.3.6 created 1976]

EC 5.4.3.7

Accepted name: leucine 2,3-aminomutase
Reaction: (2*S*)- α -leucine = (3*R*)- β -leucine
Systematic name: (2*S*)- α -leucine 2,3-aminomutase
Comments: Requires a cobamide coenzyme.
References: [83, 227, 226]

[EC 5.4.3.7 created 1982]

EC 5.4.3.8

Accepted name: glutamate-1-semialdehyde 2,1-aminomutase
Reaction: L-glutamate 1-semialdehyde = 5-aminolevulinate
Other name(s): glutamate-1-semialdehyde aminotransferase
Systematic name: (*S*)-4-amino-5-oxopentanoate 4,5-aminomutase
Comments: Requires pyridoxal phosphate.
References: [95]

[EC 5.4.3.8 created 1983]

EC 5.4.4 Transferring hydroxy groups

EC 5.4.4.1

Accepted name: (hydroxyamino)benzene mutase
Reaction: (hydroxyamino)benzene = 2-aminophenol
Other name(s): HAB mutase; hydroxylaminobenzene hydroxymutase; hydroxylaminobenzene mutase
Systematic name: (hydroxyamino)benzene hydroxymutase
References: [108, 59]

[EC 5.4.4.1 created 2003]

EC 5.4.4.2

Accepted name: isochorismate synthase
Reaction: chorismate = isochorismate
Other name(s): MenF
Systematic name: isochorismate hydroxymutase
Comments: Requires Mg²⁺. The reaction is reversible.
References: [336, 303, 55, 58]

[EC 5.4.4.2 created 1972 as EC 5.4.99.6, transferred 2003 to EC 5.4.4.2]

EC 5.4.4.3

Accepted name: 3-(hydroxyamino)phenol mutase
Reaction: 3-hydroxyaminophenol = aminohydroquinone
Other name(s): 3-hydroxylaminophenol mutase; 3HAP mutase
Systematic name: 3-(hydroxyamino)phenol hydroxymutase
References: [246]

[EC 5.4.4.3 created 2003]

EC 5.4.99 Transferring other groups

EC 5.4.99.1

Accepted name: methylaspartate mutase
Reaction: *L-threo*-3-methylaspartate = *L*-glutamate
Other name(s): glutamate mutase; glutamic mutase; glutamic isomerase; glutamic acid mutase; glutamic acid isomerase; methylaspartic acid mutase; β -methylaspartate-glutamate mutase; glutamate isomerase
Systematic name: *L-threo*-3-methylaspartate carboxy-aminomethylmutase
Comments: Requires a cobamide coenzyme.
References: [21, 309]

[EC 5.4.99.1 created 1961]

EC 5.4.99.2

Accepted name: methylmalonyl-CoA mutase
Reaction: (*R*)-methylmalonyl-CoA = succinyl-CoA
Other name(s): methylmalonyl-CoA CoA-carbonyl mutase; methylmalonyl coenzyme A mutase; methylmalonyl coenzyme A carbonylmutase; (*S*)-methylmalonyl-CoA mutase; (*R*)-2-methyl-3-oxopropanoyl-CoA CoA-carbonylmutase [incorrect]
Systematic name: (*R*)-methylmalonyl-CoA CoA-carbonylmutase
Comments: Requires a cobamide coenzyme.
References: [20]

[EC 5.4.99.2 created 1961, modified 1983]

EC 5.4.99.3

Accepted name: 2-acetolactate mutase
Reaction: 2-acetolactate = 3-hydroxy-3-methyl-2-oxobutanoate
Other name(s): acetolactate mutase; acetoxy acid isomerase
Systematic name: 2-acetolactate methylmutase
Comments: Requires ascorbic acid; also converts 2-aceto-2-hydroxybutanoate to 3-hydroxy-3-methyl-2-oxopentanoate.
References: [7]

[EC 5.4.99.3 created 1972]

EC 5.4.99.4

Accepted name: 2-methyleneglutarate mutase
Reaction: 2-methyleneglutarate = 2-methylene-3-methylsuccinate
Other name(s): α -methyleneglutarate mutase
Systematic name: 2-methyleneglutarate carboxy-methylenemethylmutase
Comments: Requires a cobamide coenzyme.
References: [155, 156]

[EC 5.4.99.4 created 1972]

EC 5.4.99.5

Accepted name: chorismate mutase
Reaction: chorismate = prephenate

Other name(s): hydroxyphenylpyruvate synthase
Systematic name: chorismate pyruvatemutase
References: [50, 169, 268, 325]

[EC 5.4.99.5 created 1972]

[5.4.99.6 *Transferred entry. isochorismate synthase. Now EC 5.4.4.2, isochorismate synthase*]

[EC 5.4.99.6 created 1972, deleted 2003]

EC 5.4.99.7

Accepted name: lanosterol synthase
Reaction: (S)-2,3-epoxysqualene = lanosterol
Other name(s): 2,3-epoxysqualene lanosterol cyclase; squalene-2,3-oxide-lanosterol cyclase; lanosterol 2,3-oxidosqualene cyclase; squalene 2,3-epoxide:lanosterol cyclase; 2,3-oxidosqualene sterol cyclase; oxidosqualene cyclase; 2,3-oxidosqualene cyclase; 2,3-oxidosqualene-lanosterol cyclase; oxidosqualene-lanosterol cyclase; squalene epoxidase-cyclase
Systematic name: (S)-2,3-epoxysqualene mutase (cyclizing, lanosterol-forming)
References: [61]

[EC 5.4.99.7 created 1961 as EC 1.99.1.13, transferred 1965 to EC 1.14.1.3, part transferred 1972 to EC 5.4.99.7 rest to EC 1.14.99.7]

EC 5.4.99.8

Accepted name: cycloartenol synthase
Reaction: (S)-2,3-epoxysqualene = cycloartenol
Other name(s): 2,3-epoxysqualene cycloartenol-cyclase; squalene-2,3-epoxide-cycloartenol cyclase; 2,3-epoxysqualene cycloartenol-cyclase; 2,3-epoxysqualene-cycloartenol cyclase; 2,3-oxidosqualene-cycloartenol cyclase
Systematic name: (S)-2,3-epoxysqualene mutase (cyclizing, cycloartenol-forming)
References: [236]

[EC 5.4.99.8 created 1972]

EC 5.4.99.9

Accepted name: UDP-galactopyranose mutase
Reaction: UDP-D-galactopyranose = UDP-D-galacto-(1→4)-furanose
Systematic name: UDP-D-galactopyranose furanomutase
References: [294]

[EC 5.4.99.9 created 1984]

[5.4.99.10 *Deleted entry. isomaltulose synthetase. Now included with EC 5.4.99.11, isomaltulose synthase*]

[EC 5.4.99.10 created 1984, deleted 1992]

EC 5.4.99.11

Accepted name: isomaltulose synthase
Reaction: sucrose = 6-O- α -D-glucopyranosyl-D-fructofuranose
Other name(s): isomaltulose synthetase; sucrose α -glucosyltransferase; trehalulose synthase
Systematic name: sucrose glucosylmutase
Comments: The enzyme simultaneously produces isomaltulose (6-O- α -D-glucopyranosyl-D-fructose) and smaller amounts of trehalulose (1-O- α -D-glucopyranosyl- β -D-fructose) from sucrose.
References: [45, 46]

[EC 5.4.99.11 created 1989 (EC 5.4.99.10 created 1984, incorporated 1992)]

EC 5.4.99.12

Accepted name: tRNA-pseudouridine synthase I
Reaction: tRNA uridine = tRNA pseudouridine
Other name(s): tRNA-uridine isomerase; tRNA pseudouridylate synthase I; transfer ribonucleate pseudouridine synthetase; pseudouridine synthase; transfer RNA pseudouridine synthetase
Systematic name: tRNA-uridine uracilmutase
Comments: The uridylate residues at positions 38, 39 and 40 of nearly all tRNAs are isomerized to pseudouridine.
References: [15, 133]

[EC 5.4.99.12 created 1990]

EC 5.4.99.13

Accepted name: isobutyryl-CoA mutase
Reaction: 2-methylpropanoyl-CoA = butanoyl-CoA
Other name(s): isobutyryl coenzyme A mutase; butyryl-CoA:isobutyryl-CoA mutase
Systematic name: 2-methylpropanoyl-CoA CoA-carbonylmutase
Comments: Requires a cobamide coenzyme.
References: [37]

[EC 5.4.99.13 created 1992]

EC 5.4.99.14

Accepted name: 4-carboxymethyl-4-methylbutenolide mutase
Reaction: 4-carboxymethyl-4-methylbut-2-en-1,4-olide = 4-carboxymethyl-3-methylbut-2-en-1,4-olide
Other name(s): 4-methyl-2-enelactone isomerase; 4-methylmuconolactone methylisomerase; 4-methyl-3-enelactone methyl isomerase
Systematic name: 4-carboxymethyl-4-methylbut-2-en-1,4-olide methylmutase
References: [40]

[EC 5.4.99.14 created 1992]

EC 5.4.99.15

Accepted name: (1→4)- α -D-glucan 1- α -D-glucosylmutase
Reaction: 4-[(1→4)- α -D-glucosyl]_{n-1}-D-glucose = 1- α -D-[(1→4)- α -D-glucosyl]_{n-1}- α -D-glucopyranoside
Other name(s): malto-oligosyltrehalose synthase; maltodextrin α -D-glucosyltransferase
Systematic name: (1→4)- α -D-glucan 1- α -D-glucosylmutase
Comments: The enzyme from *Arthrobacter sp.*, *Sulfolobus acidocaldarius* acts on (1→4)- α -D-glucans containing three or more (1→4)- α -linked D-glucose units. Not active towards maltose.
References: [183, 203, 202]

[EC 5.4.99.15 created 1999]

EC 5.4.99.16

Accepted name: maltose α -D-glucosyltransferase
Reaction: maltose = α,α -trehalose
Other name(s): trehalose synthase; maltose glucosylmutase
Systematic name: maltose α -D-glucosylmutase
References: [207, 208]

[EC 5.4.99.16 created 1999]

EC 5.4.99.17

Accepted name: squalene—hopene cyclase
Reaction: (1) squalene = hop-22(29)-ene
(2) squalene + H₂O = hopan-22-ol
Systematic name: squalene mutase (cyclizing)
Comments: The enzyme produces a constant ratio of about 5:1 hopene:hopanol.
References: [250, 117]

[EC 5.4.99.17 created 2002]

EC 5.4.99.18

Accepted name: 5-(carboxyamino)imidazole ribonucleotide mutase
Reaction: 5-carboxyamino-1-(5-phospho-D-ribosyl)imidazole = 5-amino-1-(5-phospho-D-ribosyl)imidazole-4-carboxylate
Other name(s): N⁵-CAIR mutase; PurE; N⁵-carboxyaminoimidazole ribonucleotide mutase; class I PurE
Systematic name: 5-carboxyamino-1-(5-phospho-D-ribosyl)imidazole carboxymutase
Comments: In eubacteria, fungi and plants, this enzyme, along with EC 6.3.4.18, 5-(carboxyamino)imidazole ribonucleotide synthase, is required to carry out the single reaction catalysed by EC 4.1.1.21, phosphoribosylaminoimidazole carboxylase, in vertebrates [76]. In the absence of EC 6.3.2.6, phosphoribosylaminoimidazolesuccinocarboxamide synthase, the reaction is reversible [191]. The substrate is readily converted into 5-amino-1-(5-phospho-D-ribosyl)imidazole by non-enzymic decarboxylation [191].
References: [192, 200, 191, 184, 77, 76]

[EC 5.4.99.18 created 2006]

EC 5.5 Intramolecular lyases

This subclass contains a single sub-subclass for enzymes that catalyse reactions in which a group can be regarded as being eliminated from one part of a molecule, leaving a double bond, while remaining covalently attached to the molecule (intramolecular lyases; EC 5.5.1).

EC 5.5.1 Intramolecular lyases (only sub-subclass identified to date)

EC 5.5.1.1

Accepted name: muconate cycloisomerase
Reaction: 2,5-dihydro-5-oxofuran-2-acetate = *cis,cis*-hexadienedioate
Other name(s): muconate cycloisomerase I; *cis,cis*-muconate-lactonizing enzyme; *cis,cis*-muconate cycloisomerase; muconate lactonizing enzyme; 4-carboxymethyl-4-hydroxyisocrotonolactone lyase (decyclizing); CatB; MCI
Systematic name: 2,5-dihydro-5-oxofuran-2-acetate lyase (decyclizing)
Comments: Requires Mn²⁺. Also acts (in the reverse reaction) on 3-methyl-*cis,cis*-hexadienedioate and, very slowly, on *cis,trans*-hexadienedioate. Not identical with EC 5.5.1.7 (chloromuconate cycloisomerase) or EC 5.5.1.11 (dichloromuconate cycloisomerase).
References: [214, 216, 261]

[EC 5.5.1.1 created 1961]

EC 5.5.1.2

Accepted name: 3-carboxy-*cis,cis*-muconate cycloisomerase
Reaction: 2-carboxy-2,5-dihydro-5-oxofuran-2-acetate = *cis,cis*-butadiene-1,2,4-tricarboxylate

Other name(s): β -carboxymuconate lactonizing enzyme; 3-carboxymuconolactone hydrolase
Systematic name: 2-carboxy-2,5-dihydro-5-oxofuran-2-acetate lyase (decyclizing)
References: [215, 216]

[EC 5.5.1.2 created 1972]

EC 5.5.1.3

Accepted name: tetrahydroxypteridine cycloisomerase
Reaction: tetrahydroxypteridine = xanthine-8-carboxylate
Systematic name: tetrahydroxypteridine lyase (isomerizing)
References: [188]

[EC 5.5.1.3 created 1972]

EC 5.5.1.4

Accepted name: inositol-3-phosphate synthase
Reaction: D-glucose 6-phosphate = 1D-*myo*-inositol 3-phosphate
Other name(s): *myo*-inositol-1-phosphate synthase; D-glucose 6-phosphate cycloaldolase; inositol 1-phosphate synthase; glucose 6-phosphate cyclase; inositol 1-phosphate synthetase; glucose-6-phosphate inositol monophosphate cycloaldolase; glucocycloaldolase; 1L-*myo*-inositol-1-phosphate lyase (isomerizing)
Systematic name: 1D-*myo*-inositol-3-phosphate lyase (isomerizing)
Comments: Requires NAD⁺, which dehydrogenates the -CHOH- group to -CO- at C-5 of the glucose 6-phosphate, making C-6 into an active methylene, able to condense with the -CHO at C-1. Finally, the enzyme-bound NADH reconverts C-5 into the -CHOH- form.
References: [70, 254, 22, 23]

[EC 5.5.1.4 created 1972, modified 2001]

EC 5.5.1.5

Accepted name: carboxy-*cis,cis*-muconate cyclase
Reaction: 3-carboxy-2,5-dihydro-5-oxofuran-2-acetate = 3-carboxy-*cis,cis*-muconate
Other name(s): 3-carboxymuconate cyclase
Systematic name: 3-carboxy-2,5-dihydro-5-oxofuran-2-acetate lyase (decyclizing)
References: [100]

[EC 5.5.1.5 created 1972]

EC 5.5.1.6

Accepted name: chalcone isomerase
Reaction: a chalcone = a flavanone
Other name(s): chalcone-flavanone isomerase
Systematic name: flavanone lyase (decyclizing)
References: [199]

[EC 5.5.1.6 created 1972]

EC 5.5.1.7

Accepted name: chloromuconate cycloisomerase
Reaction: 2-chloro-2,5-dihydro-5-oxofuran-2-acetate = 3-chloro-*cis,cis*-muconate
Other name(s): muconate cycloisomerase II
Systematic name: 2-chloro-2,5-dihydro-5-oxofuran-2-acetate lyase (decyclizing)

Comments: Requires Mn^{2+} . The product of cycloisomerization of 3-chloro-*cis,cis*-muconate spontaneously eliminates chloride to produce *cis*-4-carboxymethylenebut-2-en-4-olide. Also acts (in the reverse direction) on 2-chloro-*cis,cis*-muconate. Not identical with EC 5.5.1.1 (muconate cycloisomerase) or EC 5.5.1.11 (dichloromuconate cycloisomerase).

References: [247]

[EC 5.5.1.7 created 1983]

EC 5.5.1.8

Accepted name: bornyl diphosphate synthase
Reaction: geranyl diphosphate = (+)-bornyl diphosphate
Other name(s): bornyl pyrophosphate synthase; bornyl pyrophosphate synthetase; (+)-bornylpyrophosphate cyclase; geranyl-diphosphate cyclase (ambiguous)
Systematic name: (+)-bornyl-diphosphate lyase (decyclizing)
References: [53]

[EC 5.5.1.8 created 1984]

EC 5.5.1.9

Accepted name: cycloeucaenol cycloisomerase
Reaction: cycloeucaenol = obtusifoliol
Other name(s): cycloeucaenol—obtusifoliol isomerase
Systematic name: cycloeucaenol lyase (cyclopropane-decyclizing)
Comments: Opens the cyclopropane ring of a number of related 4α -methyl- 9β -19-cyclosterols, but not those with a 4β -methyl group, with formation of an 8(9) double bond. Involved in the synthesis of plant sterols.
References: [111, 230]

[EC 5.5.1.9 created 1986]

EC 5.5.1.10

Accepted name: α -pinene-oxide decyclase
Reaction: α -pinene oxide = (*Z*)-2-methyl-5-isopropylhexa-2,5-dienal
Other name(s): α -pinene oxide lyase
Systematic name: α -pinene-oxide lyase (decyclizing)
Comments: Both rings of pinene are cleaved in the reaction.
References: [98]

[EC 5.5.1.10 created 1990]

EC 5.5.1.11

Accepted name: dichloromuconate cycloisomerase
Reaction: 2,4-dichloro-2,5-dihydro-5-oxofuran-2-acetate = 2,4-dichloro-*cis,cis*-muconate
Systematic name: 2,4-dichloro-2,5-dihydro-5-oxofuran-2-acetate lyase (decyclizing)
Comments: Requires Mn^{2+} . The product of cycloisomerization of dichloro-*cis,cis*-muconate spontaneously eliminates chloride to produce *cis*-4-carboxymethylene-3-chlorobut-2-en-4-olide. Also acts, in the reverse direction, on *cis,cis*-muconate and its monochloro-derivatives, but with lower affinity. Not identical with EC 5.5.1.1 (muconate cycloisomerase) or EC 5.5.1.7 (chloromuconate cycloisomerase).
References: [154]

[EC 5.5.1.11 created 1992]

EC 5.5.1.12

Accepted name: copalyl diphosphate synthase
Reaction: geranylgeranyl diphosphate = (+)-copalyl diphosphate
Systematic name: (+)-copalyl-diphosphate lyase (deacyclizing)
Comments: Part of a bifunctional enzyme involved in the biosynthesis of abietadiene. See also EC 4.2.3.18 (abietadiene synthase)
References: [224, 225, 223, 233, 222]

[EC 5.5.1.12 created 2002]

EC 5.5.1.13

Accepted name: *ent*-copalyl diphosphate synthase
Reaction: geranylgeranyl diphosphate = *ent*-copalyl diphosphate
Other name(s): *ent*-kaurene synthase A; *ent*-kaurene synthetase A; *ent*-CDP synthase
Systematic name: *ent*-copalyl-diphosphate lyase (deacyclizing)
Comments: Part of a bifunctional enzyme involved in the biosynthesis of kaurene. See also EC 4.2.3.19 (*ent*-kaurene synthase)
References: [73, 277, 138, 292]

[EC 5.5.1.13 created 2002]

EC 5.5.1.14

Accepted name: *syn*-copalyl-diphosphate synthase
Reaction: geranylgeranyl diphosphate = 9 α -copalyl diphosphate
Other name(s): OsCyc1; OsCPSsyn; *syn*-CPP synthase; *syn*-copalyl diphosphate synthase
Systematic name: 9 α -copalyl-diphosphate lyase (deacyclizing)
Comments: Requires a divalent metal ion, preferably Mg²⁺, for activity. This class II terpene synthase produces *syn*-copalyl diphosphate, a precursor of several rice phytoalexins, including oryzalexin S and momilactones A and B. Phytoalexins are diterpenoid secondary metabolites that are involved in the defense mechanism of the plant, and are produced in response to pathogen attack through the perception of elicitor signal molecules such as chitin oligosaccharide, or after exposure to UV irradiation. The enzyme is constitutively expressed in the roots of plants where one of its products, momilactone B, acts as an allelochemical (a molecule released into the environment to suppress the growth of neighbouring plants). In other tissues the enzyme is upregulated by conditions that stimulate the biosynthesis of phytoalexins.
References: [217, 328]

[EC 5.5.1.14 created 2008]

EC 5.5.1.15

Accepted name: terpenedienyl-diphosphate synthase
Reaction: geranylgeranyl diphosphate = terpenedienyl diphosphate
Other name(s): terpenedienol diphosphate synthase; Cyc1; clerodadienyl diphosphate synthase
Systematic name: terpenedienyl-diphosphate lyase (deacyclizing)
Comments: Requires Mg²⁺. Contains a DXDD motif, which is a characteristic of diterpene cyclases whose reactions are initiated by protonation at the 14,15-double bond of geranylgeranyl diphosphate (GGDP) [103]. The triggering proton is lost at the end of the cyclization reaction [69]. The product of the reaction, terpenedienyl diphosphate, is the substrate for EC 4.2.3.36, terpenetriene synthase and is a precursor of the diterpenoid antibiotic terpentecin.
References: [57, 103, 69]

[EC 5.5.1.15 created 2008]

EC 5.5.1.16

Accepted name: halimadienyl-diphosphate synthase
Reaction: geranylgeranyl diphosphate = halima-5(6),13-dien-15-yl diphosphate
Other name(s): Rv3377c; halimadienyl diphosphate synthase; tuberculosinol diphosphate synthase
Systematic name: halima-5(6),13-dien-15-yl-diphosphate lyase (cyclizing)
Comments: Requires Mg²⁺ for activity. This enzyme is found in pathogenic prokaryotes such as *Mycobacterium tuberculosis* but not in non-pathogens such as *Mycobacterium smegmatis* so may play a role in pathogenicity. The product of the reaction is subsequently dephosphorylated yielding tuberculosinol [halima-5(6),13-dien-15-ol].
References: [206]

[EC 5.5.1.16 created 2008]

EC 5.99 Other isomerases

This subclass contains miscellaneous enzymes in a single sub-subclass (EC 5.99.1).

EC 5.99.1 Sole sub-subclass for isomerases that do not belong in the other subclasses

EC 5.99.1.1

Accepted name: thiocyanate isomerase
Reaction: benzyl isothiocyanate = benzyl thiocyanate
Other name(s): isothiocyanate isomerase
Systematic name: benzyl-thiocyanate isomerase
References: [304]

[EC 5.99.1.1 created 1965]

EC 5.99.1.2

Accepted name: DNA topoisomerase
Reaction: ATP-independent breakage of single-stranded DNA, followed by passage and rejoining
Other name(s): type I DNA topoisomerase; untwisting enzyme; relaxing enzyme; nicking-closing enzyme; swivelase; ω-protein; deoxyribonucleate topoisomerase; topoisomerase; type I DNA topoisomerase
Systematic name: DNA topoisomerase
Comments: These enzymes bring about the conversion of one topological isomer of DNA into another, *e.g.*, the relaxation of superhelical turns in DNA, the interconversion of simple and knotted rings of single-stranded DNA, and the intertwisting of single-stranded rings of complementary sequences, *cf.* EC 5.99.1.3 DNA topoisomerase (ATP-hydrolysing).
References: [89]

[EC 5.99.1.2 created 1984]

EC 5.99.1.3

Accepted name: DNA topoisomerase (ATP-hydrolysing)
Reaction: ATP-dependent breakage, passage and rejoining of double-stranded DNA
Other name(s): type II DNA topoisomerase; DNA-gyrase; deoxyribonucleate topoisomerase; deoxyribonucleic topoisomerase; topoisomerase; DNA topoisomerase II;
Systematic name: DNA topoisomerase (ATP-hydrolysing)
Comments: The enzyme can introduce negative superhelical turns into double-stranded circular DNA. One unit has nicking-closing activity, and another catalyses super-twisting and hydrolysis of ATP (*cf.* EC 5.99.1.2 DNA topoisomerase).
References: [89]

[EC 5.99.1.3 created 1984]

EC 5.99.1.4

Accepted name: 2-hydroxychromene-2-carboxylate isomerase
Reaction: 2-hydroxy-2*H*-chromene-2-carboxylate = (3*E*)-4-(2-hydroxyphenyl)-2-oxobut-3-enoate
Other name(s): HCCA isomerase; 2HC2CA isomerase; 2-hydroxychromene-2-carboxylic acid isomerase
Systematic name: 2-hydroxy-2*H*-chromene-2-carboxylate—(3*E*)-4-(2-hydroxyphenyl)-2-oxobut-3-enoate isomerase
Comments: This enzyme is involved in naphthalene degradation.
References: [212, 139, 67, 290]

[EC 5.99.1.4 created 2010]

References

- [1] D.J. Aberhart, H.-J. Lim, and B.H. Weiller. Stereochemistry of lysine 2,3-aminomutase. *J. Am. Chem. Soc.*, 103:6750–6752, 1981.
- [2] E. Adams and I.L. Norton. Purification and properties of inducible hydroxyproline 2-epimerase from *Pseudomonas*. *J. Biol. Chem.*, 239:1525–1535, 1964.
- [3] B.W. Agranoff, H. Eggerer, U. Henning, and F. Lynen. Biosynthesis of terpenes. VII. Isopentenyl pyrophosphate isomerase. *J. Biol. Chem.*, 235:326–332, 1960.
- [4] S.A. Ahmed, N. Esaki, H. Tanaka, , and K. L- α -Amino- β -thio- ϵ -caprolactam, a new sulfur-containing substrate for α -amino- ϵ -caprolactam racemase. *FEBS Lett.*, 174:76–79, 1984.
- [5] S.A. Ahmed, N. Esaki, H. Tanaka, , and K. Mechanism of α -amino- ϵ -caprolactam racemase reaction. *Biochemistry*, 25:385–388, 1986.
- [6] S. Al-Babili, P. Huguency, M. Schledz, R. Welsch, H. Frohnmeyer, O. Laule, and P. Beyer. Identification of a novel gene coding for neoxanthin synthase from *Solanum tuberosum*. *FEBS Lett.*, 485:168–172, 2000.
- [7] H.S. Allaudeen and T. Ramakrishnan. Biosynthesis of isoleucine and valine in *Mycobacterium tuberculosis* H37 Rv. *Arch. Biochem. Biophys.*, 125:199–209, 1968.
- [8] J. Altenbuchner, M. Siemann-Herzberg, and C. Sydatk. Hydantoinases and related enzymes as biocatalysts for the synthesis of unnatural chiral amino acids. *Curr. Opin. Biotechnol.*, 12:559–563, 2001.
- [9] H. Amos. A racemase for threonine in *Escherichia coli*. *J. Am. Chem. Soc.*, 76:3858–3858, 1954.
- [10] P. Anastasis, I. Freer, C. Gilmore, H. Mackie, K. Overton, D. Picken, and S. Swanson. Biosynthesis of γ -bisabolene in tissue-cultures of *Andrographis paniculata*. *Can. J. Chem.*, 62:2079–2088, 1984.
- [11] R.L. Anderson and D.P. Allison. Purification and characterization of D-lyxose isomerase. *J. Biol. Chem.*, 240:2367–2372, 1965.
- [12] A. Andersson, G. Schneider, and Y. Lindqvist. Purification and preliminary X-ray crystallographic studies of recombinant L-ribulose-5-phosphate 4-epimerase from *Escherichia coli*. *Protein Sci.*, 4:1648–1650, 1995.
- [13] R.G. Annett and G.W. Kosicki. Oxalacetate keto-enol tautomerase. Purification and characterization. *J. Biol. Chem.*, 244:2059–2067, 1969.
- [14] M. Antia, D.S. Hoare, and W. Work. The stereoisomers of $\alpha\epsilon$ -diaminopimelic acid. 3. Properties and distribution of diaminopimelic acid racemase, an enzyme causing interconversion of the LL and meso isomers. *Biochem. J.*, 65:448–459, 1957.
- [15] F. Arena, G. Ciliberto, S. Ciampi, and R. Cortese. Purification of pseudouridylate synthetase I from *Salmonella typhimurium*. *Nucleic Acids Res.*, 5:4523–4536, 1978.
- [16] G. Ashwell and J. Hickman. Enzymatic formation of xylulose 5-phosphate from ribose 5-phosphate in spleen. *J. Biol. Chem.*, 226:65–76, 1957.
- [17] G. Ashwell, A.J. Wahba, and J. Hickman. Uronic acid metabolism in bacteria. I. Purification and properties of uronic acid isomerase in *Escherichia coli*. *J. Biol. Chem.*, 235:1559–1565, 1960.
- [18] A. Baich, R.G. Wolfe, and F.J. Reithel. The enzymes of mammary gland. I. Isolation of phosphoglucose isomerase. *J. Biol. Chem.*, 235:3130–3133, 1960.
- [19] G.A. Barber and P.A. Hebda. GDP-D-mannose: GDP-L-galactose epimerase from *Chlorella pyrenoidosa*. *Methods Enzymol.*, 83:522–525, 1982.
- [20] H.A. Barker. Coenzyme B₁₂-dependent mutases causing carbon chain rearrangements. In P.D. Boyer, editor, *The Enzymes*, volume 6, pages 509–537. Academic Press, New York, 3rd edition, 1972.
- [21] H.A. Barker, V. Rooze, F. Suzuki, and A.A. Iodice. The glutamate mutase system. Assays and properties. *J. Biol. Chem.*, 239:3260–3266, 1964.

- [22] J.E.G. Barnett and D.L. Corina. The mechanism of glucose 6-phosphate-D-*myo*-inositol 1-phosphate cyclase of rat testis. The involvement of hydrogen atoms. *Biochem. J.*, 108:125–129, 1968.
- [23] J.E.G. Barnett, A. Rasheed, and D.L. Corina. Partial reactions of glucose 6-phosphate-1L-*myo*-inositol 1-phosphate cyclase. *Biochem. J.*, 131:21–30, 1973.
- [24] J.A. Beebe and P.A. Frey. Galactose mutarotase: purification, characterization, and investigations of two important histidine residues. *Biochemistry*, 37:14989–14997, 1998.
- [25] E.J. Behrman and R.Y. Stanier. The bacterial oxidation of nicotinic acid. *J. Biol. Chem.*, 228:923–945, 1957.
- [26] R. Ben-Zvi and M. Schramm. A phosphoglucomutase specific for β -glucose 1-phosphate. *J. Biol. Chem.*, 236:2186–2189, 1961.
- [27] R. Bentley and D.S. Bhate. Mutarotase from *Penicillium notatum*. I. Purification, assay, and general properties of the enzyme. *J. Biol. Chem.*, 235:1219–1224, 1960.
- [28] R. Bentley and D.S. Bhate. Mutarotase from *Penicillium notatum*. II. The mechanism of the mutarotation reaction. *J. Biol. Chem.*, 235:1225–1233, 1960.
- [29] P.S. Bernstein, W.C. Law, and R.R. Rando. Isomerization of *all-trans*-retinoids to 11-*cis*-retinoids in vitro. *Proc. Natl. Acad. Sci. USA*, 84:1849–1853, 1987.
- [30] J.M. Bishop. Cellular oncogenes and retroviruses. *Annu. Rev. Biochem.*, 52:301–354, 1983.
- [31] F. Blasi, F. Fragonmele, and I. Covelli. Thyroidal phenylpyruvate tautomerase. Isolation and characterization. *J. Biol. Chem.*, 244:4864–4870, 1969.
- [32] K. Bloch. Enzymatic synthesis of monounsaturated fatty acids. *Acc. Chem. Res.*, 2:193–202, 1969.
- [33] F. Bouvier, A. d'Harlingue, R.A. Backhaus, M.H. Kumagai, and B. Camara. Identification of neoxanthin synthase as a carotenoid cyclase paralog. *Eur. J. Biochem.*, 267:6346–6352, 2000.
- [34] F. Bouvier, P. Huguency, A. d'Harlingue, M. Kuntz, and B. Camara. Xanthophyll biosynthesis in chromoplasts: isolation and molecular cloning of an enzyme catalyzing the conversion of 5,6-epoxycarotenoid into ketocarotenoid. *Plant J.*, 6:45–54, 1994.
- [35] E. Bowman, M. McQueney, R.J. Barry, and D. Dunaway-Mariano. Catalysis and thermodynamics of the phosphoenolpyruvate phosphonopyruvate rearrangement - entry into the phosphonate class of naturally-occurring organophosphorus compounds. *J. Am. Chem. Soc.*, 110:5575–5576, 1988.
- [36] G.H. Braus, K. Luger, G. Paravicini, T. Schmidheini, K. Kirschner, and R. Hütter. The role of the TRP1 gene in yeast tryptophan biosynthesis. *J. Biol. Chem.*, 263:7868–7875, 1988.
- [37] G. Brendelberger, J. Rétey, D.M. Ashworth, K. Reynolds, F. Willenbrock, and J.A. Robinson. The enzymic interconversion of isobutyryl and *N*-butyrylcarba(dethia)-coenzyme-A - a coenzyme-B₁₂-dependent carbon skeleton rearrangement. *Angew. Chem. Int. Ed. Engl.*, 27:1089–1091, 1988.
- [38] C.D. Bridges and R.A. Alvarez. The visual cycle operates via an isomerase acting on *all-trans* retinol in the pigment epithelium. *Science*, 236:1678–1680, 1987.
- [39] D.J.H. Brock, L.R. Kass, and K. Bloch. β -Hydroxydecanoyl thioester dehydrase. II. Mode of action. *J. Biol. Chem.*, 242:4432–4440, 1967.
- [40] N.C. Bruce and R.B. Cain. β -Methylmuconolactone, a key intermediate in the dissimilation of methylaromatic compounds by a modified 3-oxoadipate pathway evolved in nocardioform actinomycetes. *FEMS Microbiol. Lett.*, 50:233–239, 1988.
- [41] F.H. Bruns, E. Noltmann, and A. Willemsen. Phosphomannose-isomerase. I. Über die Aktivitätsmessung und die Sulfhydryl-sowie die metallabhängigkeit der Enzymwirkung in einigen Tierischen Geweben. *Biochem. Z.*, 330:411–420, 1958.
- [42] D.P. Burma and B.L. Horecker. IV. γ -stereo_L/ δ -stereo_D-Ribulose-5-phosphate 4-epimerase. Pentose formation by *Lactobacillus plantarum*. *J. Biol. Chem.*, 231:1053–1064, 1958.

- [43] C. Cantwell, R. Beckmann, P. Whiteman, S.W. Queener, and E.P. Abraham. Isolation of deacetoxycephalosporin-c from fermentation broths of *Penicillium chrysogenum* transformants - construction of a new fungal biosynthetic-pathway. *Proc. R. Soc. Lond. B Biol. Sci.*, 248:283–289, 1992.
- [44] D.M. Carlson. Phosphoacetylglucosamine mutase from pig submaxillary gland. *Methods Enzymol.*, 8:179–182, 1966.
- [45] P.S.J. Cheetham. The extraction and mechanism of a novel isomaltulose-synthesizing enzyme from *Erwinia rhapontici*. *Biochem. J.*, 220:213–220, 1984.
- [46] P.S.J. Cheetham, C.E. Imber, and J. Isherwood. The formation of isomaltulose by immobilized *Erwinia rhapontici*. *Nature*, 299:628–631, 1982.
- [47] H.P. Chen, C.F. Lin, Y.J. Lee, S.S. Tsay, and S.H. Wu. Purification and properties of ornithine racemase from *Clostridium sticklandii*. *J. Bacteriol.*, 182:2052–2054, 2000.
- [48] E. Christ-Hazelhof and D.H. Nugteren. Purification and characterisation of prostaglandin endoperoxide Δ -isomerase, a cytoplasmic, glutathione-requiring enzyme. *Biochim. Biophys. Acta*, 572:43–51, 1979.
- [49] S.S. Cohen. Studies on D-ribulose and its enzymatic conversion to D-arabinose. *J. Biol. Chem.*, 201:71–84, 1953.
- [50] R.G.H. Cotton and F. Gibson. The biosynthesis of phenylalanine and tyrosine; enzymes converting chorismic acid into prephenic acid and their relationships to prephenate dehydratase and prephenate dehydrogenase. *Biochim. Biophys. Acta*, 100:76–88, 1965.
- [51] T.E. Creighton and C. Yanofsky. Chorismate to tryptophan (*Escherichia coli*) - Anthranilate synthetase, PR transferase, PRA isomerase, InGP synthetase, tryptophan synthetase. *Methods Enzymol.*, 17A:365–380, 1970.
- [52] J.E. Cronan, Rock Jr., and C.O. Biosynthesis of membrane lipids. In F.C. Neidhardt, editor, *Escherichia coli and Salmonella: Cellular and Molecular Biology*, volume 1, pages 612–636. ASM Press, Washington, DC, 2nd edition, 1996.
- [53] R. Croteau and F. Karp. Biosynthesis of monoterpenes: preliminary characterization of bornyl pyrophosphate synthetase from sage (*Salvia officinalis*) and demonstration that geranyl pyrophosphate is the preferred substrate for cyclization. *Arch. Biochem. Biophys.*, 198:512–522, 1979.
- [54] J. Crouzet, B. Cameron, L. Cauchois, S. Rigault, M.C. Rouyez, , and F. , Thibaut D., Debussche, L. Genetic and sequence analysis of an 8.7-kilobase *Pseudomonas denitrificans* fragment carrying eight genes involved in transformation of precorrin-2 to cobyrinic acid. *J. Bacteriol.*, 172:5980–5990, 1990.
- [55] C. Dahm, R. Müller, G. Schulte, K. Schmidt, and E. Leistner. The role of isochorismate hydroxymutase genes entC and menF in enterobactin and menaquinone biosynthesis in *Escherichia coli*. *Biochim. Biophys. Acta*, 1425:377–386, 1998.
- [56] K. Dahm, M. Lindlau, and H. Breuer. Steroid epimerase-a new enzyme of estrogen metabolism. *Biochim. Biophys. Acta*, 159:377–389, 1968.
- [57] T. Dairi, Y. Hamano, T. Kuzuyama, N. Itoh, K. Furihata, and H. Seto. Eubacterial diterpene cyclase genes essential for production of the isoprenoid antibiotic terpentecin. *J. Bacteriol.*, 183:6085–6094, 2001.
- [58] R. Daruwala, O. Kwon, R. Meganathan, and M.E. Hudspeth. A new isochorismate synthase specifically involved in menaquinone (vitamin K₂) biosynthesis encoded by the *menF* gene. *FEMS Microbiol. Lett.*, 140:159–163, 1996.
- [59] J.K. Davis, G.C. Paoli, Z. He, L.J. Nadeau, C.C. Somerville, and J.C. Spain. Sequence analysis and initial characterization of two isozymes of hydroxylaminobenzene mutase from *Pseudomonas pseudoalcaligenes* JS45. *Appl. Environ. Microbiol.*, 66:2965–2971, 2000.
- [60] H. de Reuse, A. Labigne, and D. Mengin-Lecreulx. The *Helicobacter pylori* ureC gene codes for a phosphoglucosamine mutase. *J. Bacteriol.*, 179:3488–3493, 1997.
- [61] P.D.G. Dean, P.R. Oritz de Montellano, K. Bloch, and E.J. Corey. A soluble 2,3-oxidosqualene sterol cyclase. *J. Biol. Chem.*, 242:3014–3015, 1967.
- [62] J.D. Deupree and W.A. Wood. L-Ribulose 5-phosphate 4-epimerase of *Aerobacter aerogenes*. Evidence for nicotinamide adenine dinucleotide-independent 4-epimerization by the crystalline enzyme. *J. Biol. Chem.*, 245:3988–3995, 1970.

- [63] D.L. DeWitt and W.L. Smith. Purification of prostacyclin synthase from bovine aorta by immunoaffinity chromatography. Evidence that the enzyme is a hemoprotein. *J. Biol. Chem.*, 258:3285–3293, 1983.
- [64] F. Dickens and D.H. Williamson. Pentose phosphate isomerase and epimerase from animal tissues. *Biochem. J.*, 64:567–578, 1956.
- [65] L. Ding, B.L. Seto, S.A. Ahmed, Coleman, and Jr. Purification and properties of the *Escherichia coli* K-12 NAD-dependent nucleotide diphosphosugar epimerase, ADP-L-glycero-D-manno-heptose 6-epimerase. *J. Biol. Chem.*, 269:24384–24390, 1994.
- [66] G.F. Domagk and R. Zech. Über den Abbau der Desoxyzucker durch Bakterienenzyme. I. L-Rhamnose-Isomerase aus *Lactobacillus plantarum*. *Biochem. Z.*, 339:145–153, 1963.
- [67] R.W. Eaton. Organization and evolution of naphthalene catabolic pathways: sequence of the DNA encoding 2-hydroxychromene-2-carboxylate isomerase and *trans-o*-hydroxybenzylidenepyruvate hydratase-aldolase from the NAH7 plasmid. *J. Bacteriol.*, 176:7757–7762, 1994.
- [68] S.W. Edwards and W.E. Knox. Homogentisate metabolism: the isomerization of maleylacetoacetate by an enzyme which requires glutathione. *J. Biol. Chem.*, 220:79–91, 1956.
- [69] T. Eguchi, Y. Dekishima, Y. Hamano, T. Dairi, H. Seto, and K. Kakinuma. A new approach for the investigation of isoprenoid biosynthesis featuring pathway switching, deuterium hyperlabeling, and ¹H NMR spectroscopy. The reaction mechanism of a novel streptomyces diterpene cyclase. *J. Org. Chem.*, 68:5433–5438, 2003.
- [70] Eisenberg and Jr. D-Myoinositol 1-phosphate as product of cyclization of glucose 6-phosphate and substrate for a specific phosphatase in rat testis. *J. Biol. Chem.*, 242:1375–1382, 1967.
- [71] E.Z. Eisenmesser, D.A. Bosco, M. Akke, and D. Kern. Enzyme dynamics during catalysis. *Science*, 295:1520–1523, 2002.
- [72] W. Ewald, H. Werbein, and I.L. Chaikoff. Evidence for the presence of 17-hydroxypregnenedione isomerase in beef adrenal cortex. *Biochim. Biophys. Acta*, 111:306–312, 1965.
- [73] R.R. Fall, , and C.A. Purification and properties of kaurene synthetase from *Fusarium moniliforme*. *J. Biol. Chem.*, 246:6913–6928, 1971.
- [74] D.S. Feingold, E.F. Neufeld, and W.Z. Hassid. The 4-epimerization and decarboxylation of uridine diphosphate D-glucuronic acid by extracts from *Phaseolus aureus* seedlings. *J. Biol. Chem.*, 235:910–913, 1960.
- [75] T. Ferenci, T. Strøm, and J.R. Quayle. Purification and properties of 3-hexulose phosphate synthase and phospho-3-hexuloisomerase from *Methylococcus capsulatus*. *Biochem. J.*, 144:477–486, 1974.
- [76] S.M. Firestine, S. Misialek, D.L. Toffaletti, T.J. Klem, J.R. Perfect, and V.J. Davisson. Biochemical role of the *Cryptococcus neoformans* ADE2 protein in fungal de novo purine biosynthesis. *Arch. Biochem. Biophys.*, 351:123–134, 1998.
- [77] S.M. Firestine, S.W. Poon, E.J. Mueller, J. Stubbe, and V.J. Davisson. Reactions catalyzed by 5-aminoimidazole ribonucleotide carboxylases from *Escherichia coli* and *Gallus gallus*: a case for divergent catalytic mechanisms. *Biochemistry*, 33:11927–11934, 1994.
- [78] G. Fischer. Peptidyl-prolyl *cis/trans* isomerases and their effectors. *Angew. Chem. Int. Ed. Engl.*, 33:1415–1436, 1994.
- [79] G. Fischer and H. Bang. The refolding of urea-denatured ribonuclease A is catalyzed by peptidyl-prolyl *cis-trans* isomerase. *Biochim. Biophys. Acta*, 828:39–42, 1985.
- [80] G. Fischer, H. Bang, and C. Mech. [Determination of enzymatic catalysis for the *cis-trans*-isomerization of peptide binding in proline-containing peptides]. *Biomed. Biochim. Acta*, 43:1101–1111, 1984.
- [81] G. Fischer, B. Wittmann-Liebold, K. Lang, T. Kiefhaber, and F.X. Schmid. Cyclophilin and peptidyl-prolyl *cis-trans* isomerase are probably identical proteins. *Nature*, 337:476–478, 1989.
- [82] V.N. Foltyn, I. Bendikov, J. De Miranda, R. Panizzutti, E. Dumin, M. Shleper, P. Li, M.D. Toney, E. Kartvelishvily, and H. Wolosker. Serine racemase modulates intracellular D-serine levels through an α,β -elimination activity. *J. Biol. Chem.*, 280:1754–1763, 2005.

- [83] I. Freer, G. Pedrocchi-Fantoni, D.J. Picken, and K.H. Overton. Stereochemistry of the leucine 2,3-aminomutase from tissue-cultures of *Andrographis paniculata*. *J. Chem. Soc. Chem. Commun.*, pages 80–82, 1981.
- [84] S. Fuchs, F. De Lorenzo, and C.B. Anfinsen. Studies on the mechanism of the enzymic catalysis of disulfide interchange in proteins. *J. Biol. Chem.*, 242:398–402, 1967.
- [85] A. Fujimoto, P. Ingram, and R.A. Smith. D-Glucose-1-phosphate:D-glucose-6-phosphotransferase. *Biochim. Biophys. Acta*, 96:91–101, 1965.
- [86] E.S. Furfine and R.H. Abeles. Intermediates in the conversion of 5'-S-methylthioadenosine to methionine in *Klebsiella pneumoniae*. *J. Biol. Chem.*, 263:9598–9606, 1988.
- [87] A. Garrido-Pertierra and R.A. Cooper. Identification and purification of distinct isomerase and decarboxylase enzymes involved in the 4-hydroxyphenylacetate pathway of *Escherichia coli*. *Eur. J. Biochem.*, 117:581–584, 1981.
- [88] R.W. Gaugler and O. Gabriel. Biological mechanisms involved in the formation of deoxy sugars. VII. Biosynthesis of 6-deoxy-L-talose. *J. Biol. Chem.*, 248:6041–6049, 1973.
- [89] M. Gellert. DNA topoisomerases. *Annu. Rev. Biochem.*, 50:879–910, 1981.
- [90] L.Y. Ghoda, T.M. Savarese, D.L. Dexter, R.E. Parks, Trackman Jr., Abeles P.C., and R.H. Characterization of a defect in the pathway for converting 5'-deoxy-5'-methylthioadenosine to methionine in a subline of a cultured heterogeneous human colon carcinoma. *J. Biol. Chem.*, 259:6715–6719, 1984.
- [91] S. Ghosh and S. Roseman. The sialic acids. IV. N-Acyl-D-glucosamine 6-phosphate 2-epimerase. *J. Biol. Chem.*, 240:1525–1530, 1965.
- [92] S. Ghosh and S. Roseman. The sialic acids. V. N-Acyl-D-glucosamine 2-epimerase. *J. Biol. Chem.*, 240:1531–1536, 1965.
- [93] L. Glaser. The biosynthesis of N-acetylgalactosamine. *J. Biol. Chem.*, 234:2801–2805, 1959.
- [94] L. Glaser. Glutamic acid racemase from *Lactobacillus arabinosus*. *J. Biol. Chem.*, 235:2095–2098, 1960.
- [95] S.P. Gough and C.G. Kannangara. Biosynthesis of δ -aminolevulinate in greening barley leaves: glutamate 1-semialdehyde aminotransferase. *Carlsberg Res. Commun.*, 43:185–194, 1978.
- [96] R.W. Gracy and E.A. Noltmann. Studies on phosphomannose isomerase. II. Characterization as a zinc metalloenzyme. *J. Biol. Chem.*, 243:4109–4116, 1968.
- [97] M. Green and S.S. Cohen. Enzymatic conversion of L-fucose to L-fuculose. *J. Biol. Chem.*, 219:557–568, 1956.
- [98] E.T. Griffiths, P.C. Harries, R. Jeffcoat, and P.W. Trudgill. Purification and properties of α -pinene oxide lyase from *Nocardia* sp. strain P18.3. *J. Bacteriol.*, 169:4980–4983, 1987.
- [99] S. Grisolia. Phosphoglyceric acid mutase. *Methods Enzymol.*, 5:236–242, 1962.
- [100] S.R. Gross, R.D. Gafford, and E.L. Tatum. The metabolism of protocatechuic acid by *Neurospora*. *J. Biol. Chem.*, 219:781–796, 1956.
- [101] C.F. Gunsalus, R.Y. Stanier, , and I.C. The enzymatic conversion of mandelic acid to benzoic acid. III. Fractionation and properties of the soluble enzymes. *J. Bacteriol.*, 66:548–553, 1953.
- [102] H., Levine Polet, , and A and. C in serum. *J. Biol. Chem.*, 250:351–357, 1975.
- [103] Y. Hamano, T. Kuzuyama, N. Itoh, K. Furihata, H. Seto, and T. Dairi. Functional analysis of eubacterial diterpene cyclases responsible for biosynthesis of a diterpene antibiotic, terpentecin. *J. Biol. Chem.*, 277:37098–37104, 2002.
- [104] M. Hamberg. Biosynthesis of 12-oxo-10,15(Z)-phytodienoic acid: identification of an allene oxide cyclase. *Biochem. Biophys. Res. Commun.*, 156:543–550, 1988.
- [105] K. Hammen-Jepersen and A. Munch-Petersen. Phosphodeoxyribomutase from *Escherichia coli*. Purification and some properties. *Eur. J. Biochem.*, 17:397–407, 1970.

- [106] R.K. Harrison and R.L. Stein. Substrate specificities of the peptidyl prolyl *cis-trans* isomerase activities of cyclophilin and FK-506 binding protein: evidence for the existence of a family of distinct enzymes. *Biochemistry*, 29:3813–3816, 1990.
- [107] S. Hartmans, J.P. Smits, M.J. van der Werf, F. Volkering, and J.A.M. de Bont. Metabolism of styrene oxide and 2-phenylethanol in the styrene-degrading *Xanthobacter* strain 124X. *Appl. Environ. Microbiol.*, 55:2850–2855, 1989.
- [108] Z. He, L.J. Nadeau, and J.C. Spain. Characterization of hydroxylaminobenzene mutase from pNBZ139 cloned from *Pseudomonas pseudoalcaligenes* JS45: a highly-associated sodium-dodecyl-sulfate-stable enzyme catalyzing an intramolecular transfer of hydroxyl group. *Eur. J. Biochem.*, 267:1110–1116, 2000.
- [109] E.C. Heath, B.L. Horecker, P.Z. Smyrniotis, and Y. Takagi. Pentose formation by *Lactobacillus plantarum*. II. L-Arabinose isomerase. *J. Biol. Chem.*, 231:1031–1037, 1958.
- [110] P.A. Hebda, E.J. Behrman, and G.A. Barber. The guanosine 5'-diphosphate D-mannose: guanosine 5'-diphosphate L-galactose epimerase of *Chlorella pyrenoidosa*. Chemical synthesis of guanosine 5'-diphosphate L-galactose and further studies of the enzyme and the reaction it catalyzes. *Arch. Biochem. Biophys.*, 194:496–502, 1979.
- [111] R. Heintz and P. Benveniste. Plant sterol metabolism. Enzymatic cleavage of the 9 β ,19 β -cyclopropane ring of cyclopropyl sterols in bramble tissue cultures. *J. Biol. Chem.*, 249:4267–4274, 1974.
- [112] L. Hennig, C. Christner, M. Kipping, B. Schelbert, K.P. Rucknagel, S. Grabley, G. Kullertz, and G. Fischer. Selective inactivation of parvulin-like peptidyl-prolyl *cis/trans* isomerases by juglone. *Biochemistry*, 37:5953–5960, 1998.
- [113] T. Hikada, S. Imai, O. Hara, H. Anzai, T. Murakami, K. Nagaoka, and H. Seto. Carboxyphosphoenolpyruvate phosphonmutase, a novel enzyme catalyzing C-P bond formation. *J. Bacteriol.*, 172:3066–3072, 1990.
- [114] R.M. Hochster and R.W. Watson. Enzymatic isomerization of D-xylose to D-xylulose. *Arch. Biochem. Biophys.*, 48:120–129, 1954.
- [115] B.L. Horecker, P.Z. Smyrniotis, and J.E. Seegmiller. The enzymatic conversion of 6-phosphogluconate to ribulose-5-phosphate and ribose-5-phosphate. *J. Biol. Chem.*, 193:383–396, 1951.
- [116] S. Hoshiko, Y. Kunimoto, K. Arima, and T. Beppu. Mechanism of L-alloisocitric acid fermentation: isocitrate epimerase activity in the cell-free-extract of *Penicillium purpurogenum*. *Agric. Biol. Chem.*, 46:143–151, 1982.
- [117] T. Hoshino and T. Sato. Squalene-hopene cyclase: catalytic mechanism and substrate recognition. *Chem. Commun.*, pages 291–301, 2002.
- [118] H.T. Huang. *dl*-Lysine production by lysine racemase, 1960.
- [119] R. Hubbard. Retinene isomerase. *J. Gen. Physiol.*, 39:935–962, 1956.
- [120] F.M. Huennekens, H.R. Mahler, and J. Nordmann. Studies on the cyclophorase system. XVII. The occurrence and properties of an α -hydroxy acid racemase. *Arch. Biochem.*, 30:77–89, 1951.
- [121] J. Hurwitz and B.L. Horecker. The purification of phosphoketopentoepimerase from *Lactobacillus pentosus* and the preparation of xylulose 5-phosphate. *J. Biol. Chem.*, 223:993–1008, 1956.
- [122] J. Hurwitz, A. Weissbach, B.L. Horecker, and P.Z. Smyrniotis. Spinach phosphoribulokinase. *J. Biol. Chem.*, 218:769–783, 1956.
- [123] R. Hütter, P. Niederberger, and J.A. DeMoss. Tryptophan synthetic genes in eukaryotic microorganisms. *Annu. Rev. Microbiol.*, 40:55–77, 1986.
- [124] K. Izumori, A.W. Rees, and A.D. Elbein. Purification, crystallization, and properties of D-ribose isomerase from *Mycobacterium smegmatis*. *J. Biol. Chem.*, 250:8085–8087, 1975.
- [125] B. Jacobson and E.A. Davidson. Biosynthesis of uronic acids by skin enzymes. I. Uridine diphosphate-D-glucuronic acid-5-epimerase. *J. Biol. Chem.*, 237:638–642, 1962.
- [126] I. Jacobsson, G. Bäckström, M. Höök, U. Lindahl, D.S. Feingold, A. Malmström, and L. Rodén. Biosynthesis of heparin. Assay and properties of the microsomal uronosyl C-5 epimerase. *J. Biol. Chem.*, 254:2975–2982, 1979.

- [127] L. Jolly, P. Ferrari, D. Blanot, J. van Heijenoort, F. Fassy, and D. Mengin-Lecreulx. Reaction mechanism of phosphoglucosamine mutase from *Escherichia coli*. *Eur. J. Biochem.*, 262:202–210, 1999.
- [128] L. Jolly, F. Pompeo, J. van Heijenoort, F. Fassy, and D. Mengin-Lecreulx. Autophosphorylation of phosphoglucosamine mutase from *Escherichia coli*. *J. Bacteriol.*, 182:1280–1285, 2000.
- [129] L. Jolly, S. Wu, J. van Heijenoort, H. de Lencastre, D. Mengin-Lecreulx, and A. Tomas. The femR315 gene from *Staphylococcus aureus*, the interruption of which results in reduced methicillin resistance, encodes a phosphoglucosamine mutase. *J. Bacteriol.*, 179:5321–5325, 1997.
- [130] J.G. Joshi and P. Handler. Phosphoglucomutase. I. Purification and properties of phosphoglucomutase from *Escherichia coli*. *J. Biol. Chem.*, 239:2741–2751, 1964.
- [131] R.E. Kallio and A.D. Larson. Methionine degradation by a species of *Pseudomonas*. In W.D. McElroy and H.B. Glass, editors, *A Symposium on Amino Acid Metabolism*, pages 616–634. Johns Hopkins Press, Baltimore, 1955.
- [132] H.O. Kammen and R. Koo. Phosphopentomutases. I. Identification of two activities in rabbit tissues. *J. Biol. Chem.*, 244:4888–4893, 1969.
- [133] H.O. Kammen, C.C. Marvel, L. Hardy, and E.E. Penhoet. Purification, structure, and properties of *Escherichia coli* tRNA pseudouridine synthase I. *J. Biol. Chem.*, 263:2255–2263, 1988.
- [134] K. Kaneda, T. Kuzuyama, M. Takagi, Y. Hayakawa, and H. Seto. An unusual isopentenyl diphosphate isomerase found in the mevalonate pathway gene cluster from *Streptomyces* sp. strain CL190. *Proc. Natl. Acad. Sci. USA*, 98:932–937, 2001.
- [135] V. Sri Kannathasan, A.G. Staines, C.J. Dong, R.A. Field, A.G. Preston, D.J. Maskell, and J.H. Naismith. Overexpression, purification, crystallization and data collection on the *Bordetella pertussis* *wlbD* gene product, a putative UDP-GlcNAc 2'-epimerase. *Acta Crystallogr. D Biol. Crystallogr.*, 57:1310–1312, 2001.
- [136] N. Kato, H. Yurimoto, and R.K. Thauer. The physiological role of the ribulose monophosphate pathway in bacteria and archaea. *Biosci. Biotechnol. Biochem.*, 70:10–21, 2006.
- [137] F.S. Kawahara and P. Talalay. Crystalline Δ^5 -3-ketosteroid isomerase. *J. Biol. Chem.*, 235:PC1–PC2, 1960.
- [138] H. Kawaide, R. Imai, T. Sassa, and Y. Kamiya. *Ent*-kaurene synthase from the fungus *Phaeosphaeria* sp. L487. cDNA isolation, characterization, and bacterial expression of a bifunctional diterpene cyclase in fungal gibberellin biosynthesis. *J. Biol. Chem.*, 272:21706–21712, 1997.
- [139] A. Keck, D. Conradt, A. Mahler, A. Stolz, R. Mattes, and J. Klein. Identification and functional analysis of the genes for naphthalenesulfonate catabolism by *Sphingomonas xenophaga* BN6. *Microbiology*, 152:1929–1940, 2006.
- [140] D. Keilin and E.F. Hartree. Biological catalysis of mutarotation of glucose. *Biochem. J.*, 50:341–348, 1952.
- [141] C.R. Kepler and S.B. Tove. Biohydrogenation of unsaturated fatty acids. III. Purification and properties of linoleate Δ^{12} -*cis*, Δ^{11} -*trans*-isomerase from *Butyrivibrio fibrosolvens*. *J. Biol. Chem.*, 242:5686–5692, 1967.
- [142] K. Kikuchi and S. Tsuiki. Purification and properties of UDP-*N*-acetylglucosamine 2'-epimerase from rat liver. *Biochim. Biophys. Acta*, 327:193–206, 1973.
- [143] W.W. Kilgore and M.P. Starr. Catabolism of galacturonic and glucuronic acids by *Erwinia carotovora*. *J. Biol. Chem.*, 234:2227–2235, 1959.
- [144] M.S. Kim and D.H. Shin. A preliminary X-ray study of sedoheptulose-7-phosphate isomerase from *Burkholderia pseudomallei*. *Acta Crystallogr. Sect. F Struct. Biol. Cryst. Commun.*, 65:1110–1112, 2009.
- [145] K. Kitahara, A. Obayashi, and S. Fukui. Racemase I cell-free racemase. *Enzymologia*, 15:259–266, 1953.
- [146] R.B. Kjonaas, K.V. Venkatachalam, and R. Croteau. Metabolism of monoterpenes: oxidation of isopiperitenol to isopiperitenone, and subsequent isomerization to piperitenone by soluble enzyme preparations from peppermint (*Mentha piperita*) leaves. *Arch. Biochem. Biophys.*, 238:49–60, 1985.
- [147] J.P. Klinman and I.A. Rose. Mechanism of the aconitate isomerase reaction. *Biochemistry*, 10:2259–2266, 1971.

- [148] J.P. Klinman and I.A. Rose. Purification and kinetic properties of aconitate isomerase from *Pseudomonas putida*. *Biochemistry*, 10:2253–2259, 1971.
- [149] B. Kneidinger, M. Graninger, M. Puchberger, P. Kosma, and P. Messner. Biosynthesis of nucleotide-activated D-glycero-D-manno-heptose. *J. Biol. Chem.*, 276:20935–20944, 2001.
- [150] B. Kneidinger, C. Marolda, M. Graninger, A. Zamyatina, F. McArthur, P. Kosma, M.A. Valvano, and P. Messner. Biosynthesis pathway of ADP-L-glycero- β -D-manno-heptose in *Escherichia coli*. *J. Bacteriol.*, 184:363–369, 2002.
- [151] W.E. Knox. *p*-Hydroxyphenylpyruvate enol-keto tautomerase. *Methods Enzymol.*, 2:289–295, 1955.
- [152] W.E. Knox and B.M. Pitt. Enzymic catalysis of the keto-enol tautomerization of phenylpyruvic acids. *J. Biol. Chem.*, 225:675–688, 1957.
- [153] S. Kornfeld and L. Glaser. The synthesis of thymidine-linked sugars. V. Thymidine diphosphate-amino sugars. *J. Biol. Chem.*, 237:3052–3059, 1962.
- [154] A.E. Kuhm, M. Schlömann, H.-J. Knackmuss, and D.H. Pieper. Purification and characterization of dichloromuconate cycloisomerase from *Alcaligenes eutrophus* JMP 134. *Biochem. J.*, 266:877–883, 1990.
- [155] H.-F. Kung, S. Cederbaum, L. Tsai, and T.C. Stadtman. Nicotinic acid metabolism. V. A cobamide coenzyme-dependent conversion of α -methyleneglutaric acid to dimethylmaleic acid. *Proc. Natl. Acad. Sci. USA*, 65:978–984, 1970.
- [156] H.-F. Kung and T.C. Stadtman. Nicotinic acid metabolism. VI. Purification and properties of α -methyleneglutarate mutase (B_{12} -dependent) and methylitaconate isomerase. *J. Biol. Chem.*, 246:3378–3388, 1971.
- [157] Z. Kurylo-Borowska and T. Abramsky. Biosynthesis of β -tyrosine. *Biochim. Biophys. Acta*, 264:1–10, 1972.
- [158] L. Lack. Enzymic *cis-trans* isomerization of maleylpyruvic acid. *J. Biol. Chem.*, 236:2835–2840, 1961.
- [159] L. Laiz, P. Liras, J.M. Castro, and J.F. Martín. Purification and characterization of the isopenicillin-N epimerase from *Nocardia lactamdurans*. *J. Gen. Microbiol.*, 136:663–671, 1990.
- [160] H.C. Lamont, W.L. Staudenbauer, and J.L. Strominger. Partial purification and characterization of an aspartate racemase from *Streptococcus faecalis*. *J. Biol. Chem.*, 247:5103–5106, 1972.
- [161] L.V. Lee, R.R. Poyner, M.V. Vu, and W.W. Cleland. Role of metal ions in the reaction catalyzed by L-ribulose-5-phosphate 4-epimerase. *Biochemistry*, 39:4821–4830, 2000.
- [162] N. Lee, J.W. Patrick, and M. Masson. Crystalline L-ribulose 5-phosphate 4-epimerase from *Escherichia coli*. *J. Biol. Chem.*, 243:4700–4705, 1968.
- [163] V. Lefebvre, M. Kuntz, B. Camara, and A. Palloix. The capsanthin-capsorubin synthase gene: a candidate gene for the *y* locus controlling the red fruit colour in pepper. *Plant Mol. Biol.*, 36:785–789, 1998.
- [164] L.F. Leloir. Enzymic isomerization and related processes. *Adv. Enzymol. Relat. Subj. Biochem.*, 14:193–218, 1953.
- [165] L.F. Leloir and C.E. Cardini. Enzymes acting on glucosamine phosphates. *Biochim. Biophys. Acta*, 20:33–42, 1956.
- [166] G.B. Levy and E.S. Cook. A rotographic study of mutarotase. *Biochem. J.*, 57:50–55, 1954.
- [167] H.-W. Liu and J.S. Thorson. Pathways and mechanisms in the biogenesis of novel deoxysugars by bacteria. *Annu. Rev. Microbiol.*, 48:223–256, 1994.
- [168] L. Liu, K. Iwata, A. Kita, Y. Kawarabayasi, M. Yohda, and K. Miki. Crystal structure of aspartate racemase from *Pyrococcus horikoshii* OT3 and its implications for molecular mechanism of PLP-independent racemization. *J. Mol. Biol.*, 319:479–489, 2002.
- [169] J.H. Lorence and E.W. Nester. Multiple molecular forms of chorismate mutase in *Bacillus subtilis*. *Biochemistry*, 6:1541–1543, 1967.
- [170] F. De Lorenzo, R.F. Goldberger, E. Steers, D. Givol, and C.B. Anfinsen. Purification and properties of an enzyme from beef liver which catalyzes sulfhydryl-disulfide interchange in proteins. *J. Biol. Chem.*, 241:1562–1567, 1966.

- [171] Z. Lu, , and E.C.C. The nucleotide sequence of *Escherichia coli* genes for L-fucose dissimilation. *Nucleic Acids Res.*, 17:4883–4884, 1989.
- [172] F. Lynen, J. Knappe, E. Lorch, G. Jütting, and E. Ringelmann. Die biochemische Funktion des Biotins. *Angew. Chem.*, 71:481–486, 1959.
- [173] F. Maley and G.F. Maley. The enzymic conversion of glucosamine to galactosamine. *Biochim. Biophys. Acta*, 31:577–578, 1959.
- [174] A. Malmström and L. Åberg. Biosynthesis of dermatan sulphate. Assay and properties of the uronosyl C-5 epimerase. *Biochem. J.*, 201:489–493, 1982.
- [175] M.N. Margolies and R.F. Goldberger. Isolation of the fourth (isomerase) of histidine biosynthesis from *Salmonella typhimurium*. *J. Biol. Chem.*, 241:3262–3269, 1966.
- [176] A.G. Marr and P.W. Wilson. The alanine racemase of *Brucella abortus*. *Arch. Biochem. Biophys.*, 49:424–433, 1954.
- [177] H. Marrakchi, K.H. Choi, and C.O. Rock. A new mechanism for anaerobic unsaturated fatty acid formation in *Streptococcus pneumoniae*. *J. Biol. Chem.*, 277:44809–44816, 2002.
- [178] K.O. Martin, S.-W. Oh, H.J. Lee, and C. Monder. Studies on 21-³H-labeled corticosteroids: evidence for isomerization of the ketol side chain of 11-deoxycorticosterone by a hamster liver enzyme. *Biochemistry*, 16:3803–3809, 1977.
- [179] L.A. Martinez-Cruz, M.K. Dreyer, D.C. Boisvert, H. Yokota, M.L. Martinez-Chantar, R. Kim, and S.H. Kim. Crystal structure of MJ1247 protein from *M. jannaschii* at 2.0 Å resolution infers a molecular function of 3-hexulose-6-phosphate isomerase. *Structure*, 10:195–204, 2002.
- [180] S. Martínez-Rodríguez, M. Andújar-Sánchez, J.L. Neira, J.M. Clemente-Jiménez, V. Jara-Pérez, F. Rodríguez-Vico, and F.J. Las Heras-Vázquez. Site-directed mutagenesis indicates an important role of cysteines 76 and 181 in the catalysis of hydantoin racemase from *Sinorhizobium meliloti*. *Protein Sci.*, 15:2729–2738, 2006.
- [181] S. Martínez-Rodríguez, F.J. Las Heras-Vázquez, J.M. Clemente-Jiménez, and F. Rodríguez-Vico. Biochemical characterization of a novel hydantoin racemase from *Agrobacterium tumefaciens* C58. *Biochimie*, 86:77–81, 2004.
- [182] S. Martínez-Rodríguez, F.J. Las Heras-Vázquez, L. Mingorance-Cazorla, J.M. Clemente-Jiménez, and F. Rodríguez-Vico. Molecular cloning, purification, and biochemical characterization of hydantoin racemase from the legume symbiont *Sinorhizobium meliloti* CECT 4114. *Appl. Environ. Microbiol.*, 70:625–630, 2004.
- [183] K. Maruta, T. Nakada, M. Kubota, H. Chaen, T. Sugimoto, M. Kurimoto, , and Y. Formation of trehalose from maltooligosaccharides by a novel enzymatic system. *Biosci. Biotechnol. Biochem.*, 59:1829–1834, 1995.
- [184] I.I. Mathews, T.J. Kappock, J. Stubbe, and S.E. Ealick. Crystal structure of *Escherichia coli* PurE, an unusual mutase in the purine biosynthetic pathway. *Structure*, 7:1395–1406, 1999.
- [185] S. Matsushashi. Enzymatic synthesis of cytidine diphosphate 3,6-dideoxyhexoses. II. Reversible 2-epimerization of cytidine diphosphate paratose. *J. Biol. Chem.*, 241:4275–4282, 1966.
- [186] E.S. Maxwell and H. de Robichon-Szulmajster. Purification of uridine diphosphate galactose-4-epimerase from yeast and the identification of protein-bound diphosphopyridine nucleotide. *J. Biol. Chem.*, 235:308–312, 1960.
- [187] R. Mazumder, T. Sasakawa, Y. Kaziro, and S. Ochoa. Metabolism of propionic acid in animal tissues. IX. Methylmalonyl coenzyme A racemase. *J. Biol. Chem.*, 237:3065–3068, 1962.
- [188] W.S. McNutt and S.P. Damle. Tetraoxypteridine isomerase. *J. Biol. Chem.*, 239:4272–4279, 1964.
- [189] A. Melo and L. Glaser. The mechanism of 6-deoxyhexose synthesis. II. Conversion of deoxythymidine diphosphate 4-keto-6-deoxy-D-glucose to deoxythymidine diphosphate L-rhamnose. *J. Biol. Chem.*, 243:1475–1478, 1968.
- [190] D. Mengin-Lecreulx and J. van Heijenoort. Characterization of the essential gene glmM encoding phosphoglucosamine mutase in *Escherichia coli*. *J. Biol. Chem.*, 271:32–39, 1996.
- [191] E. Meyer, T.J. Kappock, C. Osuji, and J. Stubbe. Evidence for the direct transfer of the carboxylate of N⁵-carboxyaminoimidazole ribonucleotide (N⁵-CAIR) to generate 4-carboxy-5-aminoimidazole ribonucleotide catalyzed by *Escherichia coli* PurE, an N⁵-CAIR mutase. *Biochemistry*, 38:3012–3018, 1999.

- [192] E. Meyer, N.J. Leonard, B. Bhat, J. Stubbe, and J.M. Smith. Purification and characterization of the *purE*, *purK*, and *purC* gene products: identification of a previously unrecognized energy requirement in the purine biosynthetic pathway. *Biochemistry*, 31:5022–5032, 1992.
- [193] E. Meyer-Arendt, G. Beisenherz, and T. Bücher. Triosephosphate isomerase. *Naturwissenschaften*, 40:59–59, 1953.
- [194] O. Meyerhof and L.V. Beck. Triosephosphate isomerase. *J. Biol. Chem.*, 156:109–120, 1944.
- [195] F.M. Miesowicz and K. Bloch. Purification of hog liver isomerase. Mechanism of isomerization of 3-alkenyl and 3-alkynyl thioesters. *J. Biol. Chem.*, 254:5868–5877, 1979.
- [196] J. De Miranda, A. Santoro, S. Engelender, and H. Wolosker. Human serine racemase: molecular cloning, genomic organization and functional analysis. *Gene*, 256:183–188, 2000.
- [197] C. Monder, K.O. Martin, and J. Bogumil. Presence of epimerase activity in hamster liver corticosteroid side chain isomerase. *J. Biol. Chem.*, 255:7192–7198, 1980.
- [198] C.G.D. Morley and T.C. Stadtman. Studies on the fermentation of D- α -lysine. Purification and properties of an adenosine triphosphate regulated B₁₂-coenzyme-dependent D- α -lysine mutase complex from *Clostridium sticklandii*. *Biochemistry*, 9:4890–4900, 1970.
- [199] E. Moustafa and E. Wong. Purification and properties of chalcone-flavanone isomerase from soya bean seed. *Phytochemistry*, 6:625–632, 1967.
- [200] E.J. Mueller, E. Meyer, J. Rudolph, V.J. Davisson, and J. Stubbe. N⁵-Carboxyaminoimidazole ribonucleotide: evidence for a new intermediate and two new enzymatic activities in the de novo purine biosynthetic pathway of *Escherichia coli*. *Biochemistry*, 33:2269–2278, 1994.
- [201] V.A. Najjar. Phosphoglucomutase. In P.D. Boyer, H. Lardy, and K. Myrbäck, editors, *The Enzymes*, volume 6, pages 161–178. Academic Press, New York, 2nd edition, 1962.
- [202] T. Nakada, S. Ikegami, H. Chaen, M. Kubota, S. Fukuda, T. Sugimoto, M. Kurimoto, , and Y. Purification and characterization of thermostable maltooligosyl trehalose synthase from the thermoacidophilic archaeobacterium *Sulfolobus acidocaldarius*. *Biosci. Biotechnol. Biochem.*, 60:263–266, 1996.
- [203] T. Nakada, K. Maruta, K. Tsusaki, M. Kubota, H. Chaen, T. Sugimoto, M. Kurimoto, , and Y. Purification and properties of a novel enzyme, maltooligosyl trehalose synthase, from *Arthrobacter* sp. Q36. *Biosci. Biotechnol. Biochem.*, 59:2210–2214, 1995.
- [204] Y. Nakagawa and E.A. Noltmann. Isolation of crystalline phosphoglucose isomerase from brewers' yeast. *J. Biol. Chem.*, 240:1877–1881, 1965.
- [205] T. Nakamatu and K. Yamanaka. Crystallization and properties of L-arabinose isomerase from *Lactobacillus gayonii*. *Biochim. Biophys. Acta*, 178:156–165, 1969.
- [206] C. Nakano, T. Okamura, T. Sato, T. Dairi, and T. Hoshino. *Mycobacterium tuberculosis* H37Rv3377c encodes the diterpene cyclase for producing the halimane skeleton. *Chem. Commun. (Camb.)*, pages 1016–1018, 2005.
- [207] T. Nishimoto, M. Nakano, S. Ikegami, H. Chaen, S. Fukuda, T. Sugimoto, M. Kurimoto, , and Y. Existence of a novel enzyme converting maltose to trehalose. *Biosci. Biotechnol. Biochem.*, 59:2189–2190, 1995.
- [208] T. Nishimoto, M. Nakano, T. Nakada, H. Chaen, S. Fukuda, T. Sugimoto, M. Kurimoto, , and Y. Purification and properties of a novel enzyme, trehalose synthase, from *Pimelobacter* sp. R48. *Biosci. Biotechnol. Biochem.*, 60:640–644, 1996.
- [209] E. Noltmann and F.H. Bruns. Reindarstellung und Eigenschaften von Phosphoglucose-isomerase aus Hefe. *Biochem. Z.*, 331:436–445, 1959.
- [210] E.A. Noltmann. Isolation of crystalline phosphoglucose isomerase from rabbit muscle. *J. Biol. Chem.*, 239:1545–1550, 1964.
- [211] N. Ogino, T. Miyamoto, S. Yamamoto, and O. Hayaishi. Prostaglandin endoperoxide E isomerase from bovine vesicular gland microsomes, a glutathione-requiring enzyme. *J. Biol. Chem.*, 252:890–895, 1977.

- [212] T. Ohmoto, T. Kinoshita, K. Moriyoshi, K. Sakai, N. Hamada, and T. Ohe. Purification and some properties of 2-hydroxychromene-2-carboxylate isomerase from naphthalenesulfonate-assimilating *Pseudomonas* sp. TA-2. *J. Biochem.*, 124:591–597, 1998.
- [213] I. Orita, H. Yurimoto, R. Hirai, Y. Kawarabayasi, Y. Sakai, and N. Kato. The archaeon *Pyrococcus horikoshii* possesses a bifunctional enzyme for formaldehyde fixation via the ribulose monophosphate pathway. *J. Bacteriol.*, 187:3636–3642, 2005.
- [214] L.N. Ornston. The conversion of catechol and protocatechuate to β -keto adipate by *Pseudomonas putida*. 3. Enzymes of the catechol pathway. *J. Biol. Chem.*, 241:3795–3799, 1966.
- [215] L.N. Ornston. The conversion of catechol and protocatechuate to β -keto adipate by *Pseudomonas putida*. II. Enzymes of the protocatechuate pathway. *J. Biol. Chem.*, 241:3787–3794, 1966.
- [216] L.N. Ornston. Conversion of catechol and protocatechuate to β -keto adipate (*Pseudomonas putida*). *Methods Enzymol.*, 17A:529–549, 1970.
- [217] K. Otomo, H. Kenmoku, H. Oikawa, W.A. König, H. Toshima, W. Mitsunashi, H. Yamane, T. Sassa, and T. Toyomasu. Biological functions of *ent*- and *syn*-copalyl diphosphate synthases in rice: key enzymes for the branch point of gibberellin and phytoalexin biosynthesis. *Plant J.*, 39:886–893, 2004.
- [218] P. Overath, G.M. Kellerman, F. Lynen, H.P. Fritz, and H.J. Keller. Zum Mechanismus der Umlagerung von Methylmalonyl-CoA in Succinyl-CoA. II. Versuche zur Wirkungsweise von Methylmalonyl-CoA-Isomerase und Methylmalonyl-CoA-Racemase. *Biochem. Z.*, 335:500–518, 1962.
- [219] N.J. Palleroni and M. Doudoroff. Mannose isomerase of *Pseudomonas saccharophila*. *J. Biol. Chem.*, 218:535–548, 1956.
- [220] J.M. Pawelek. Dopachrome conversion factor functions as an isomerase. *Biochem. Biophys. Res. Commun.*, 166:1328–1333, 1990.
- [221] J.L. Pennock, J.M. Behnke, Q.D. Bickle, E. Devaney, R.K. Grecis, , and R.E. , Joshua. G.W., Selkirk. M.E., Zhang. Y. and Meyer, D.J. Rapid purification and characterization of L-dopachrome-methyl ester tautomerase (macrophage-migration-inhibitory factor) from *Trichinella spiralis*, *Trichuris muris* and *Brugia pahangi*. *Biochem. J.*, 335:495–498, 1998.
- [222] R.J. Peters and R.B. Croteau. Abietadiene synthase catalysis: conserved residues involved in protonation-initiated cyclization of geranylgeranyl diphosphate to (+)-copalyl diphosphate. *Biochemistry*, 41:1836–1842, 2002.
- [223] R.J. Peters and R.B. Croteau. Abietadiene synthase catalysis: mutational analysis of a prenyl diphosphate ionization-initiated cyclization and rearrangement. *Proc. Natl. Acad. Sci. USA*, 99:580–584, 2002.
- [224] R.J. Peters, J.E. Flory, R. Jetter, M.M. Ravn, H.J. Lee, R.M. Coates, and R.B. Croteau. Abietadiene synthase from grand fir (*Abies grandis*): characterization and mechanism of action of the "pseudomature" recombinant enzyme. *Biochemistry*, 39:15592–15602, 2000.
- [225] R.J. Peters, M.M. Ravn, R.M. Coates, and R.B. Croteau. Bifunctional abietadiene synthase: free diffusive transfer of the (+)-copalyl diphosphate intermediate between two distinct active sites. *J. Am. Chem. Soc.*, 123:8974–8978, 2001.
- [226] J.M. Poston. Coenzyme B₁₂-dependent enzymes in potatoes: leucine 2,3-aminomutase and methylmalonyl-CoA mutase. *Phytochemistry*, 17:401–402, 1976.
- [227] J.M. Poston. Leucine 2,3-aminomutase, an enzyme of leucine catabolism. *J. Biol. Chem.*, 251:1859–1863, 1976.
- [228] J. Preiss. 4-Deoxy-L-threo-5-hexosulose uronic acid isomerase. *Methods Enzymol.*, 9:602–604, 1966.
- [229] C.R.H. Raetz. Biochemistry of endotoxins. *Annu. Rev. Biochem.*, 58:129–170, 1990.
- [230] A. Rahier, P. Schmitt, and P. Benveniste. 7-oxo-24 ξ (28)-dihydrocycloeucaleanol, a potent inhibitor of plant sterol biosynthesis. *Phytochemistry*, 21:1969–1974, 1982.
- [231] T. Ramasarma and K.V. Giri. Phosphoglucose isomerase of green gram (*Phaseolus radiatus*). *Arch. Biochem. Biophys.*, 62:91–96, 1956.
- [232] S. Ranjan, K.K. Patnaik, and M.M. Laloraya. Enzymic conversion of *meso*-tartrate to *dextro*-tartrate in tamarind. *Naturwissenschaften*, 48:406–406, 1961.

- [233] M.M. Ravn, R.J. Peters, R.M. Coates, and R. Croteau. Mechanism of abietadiene synthase catalysis: stereochemistry and stabilization of the cryptic pimarenyl carbocation intermediates. *J. Am. Chem. Soc.*, 124:6998–7006, 2002.
- [234] W.J. Ray, Jr., Peck, and Jr. Phosphomutases. In P.D. Boyer, editor, *The Enzymes*, volume 6, pages 407–477. 3rd edition, 1972.
- [235] W.J. Ray and G.A. Roscelli. A kinetic study of the phosphoglucomutase pathway. *J. Biol. Chem.*, 239:1228–1236, 1964.
- [236] H.H. Rees, L.J. Goad, and T.W. Goodwin. 2,3-Oxidosqualene cycloartenol cyclase from *Ochromonas malhamensis*. *Biochim. Biophys. Acta*, 176:892–894, 1969.
- [237] J.L. Reissig and L.F. Leloir. Phosphoacetylglucosamine mutase from *Neurospora*. *Methods Enzymol.*, 8:175–178, 1966.
- [238] J. Retey, F. Kunz, D. Arigoni, and T.C. Stadtman. Zur Kenntnis der β -Lysin-Mutase-Reaktion: mechanismus und sterischer Verlauf. *Helv. Chim. Acta*, 61:2989–2998, 1978.
- [239] H.C. Rilling and M.J. Coon. The enzymatic isomerization of α -methylvinylacetyl coenzyme A and the specificity of a bacterial α -methylcrotonyl coenzyme A carboxylase. *J. Biol. Chem.*, 235:3087–3092, 1960.
- [240] C.A. Roessner, M.J. Warren, P.J. Santander, B.P. Atshaves, S. Ozaki, N.J. Stolowich, K. Iida, , and A.I. Expression of *Salmonella typhimurium* enzymes for cobinamide synthesis. Identification of the 11-methyl and 20-methyl transferases of corrin biosynthesis. *FEBS Lett.*, 301:73–78, 1992.
- [241] R.J. Van Rooijen, S. Van Schalkwijk, , and W.M. Molecular cloning, characterization, and nucleotide sequence of the tagatose 6-phosphate pathway gene cluster of the lactose operon of *Lactococcus lactis*. *J. Biol. Chem.*, 266:7176–7181, 1991.
- [242] Z.B. Rose. The purification and properties of diphosphoglycerate mutase from human erythrocytes. *J. Biol. Chem.*, 243:4810–4820, 1968.
- [243] Z.B. Rose. The enzymology of 2,3-bisphosphoglycerate. *Adv. Enzymol. Relat. Areas Mol. Biol.*, 51:211–253, 1980.
- [244] J.R. Roth, J.G. Lawrence, M. Rubenfield, S. Kieffer-Higgins, , and G.M. Characterization of the cobalamin (vitamin B₁₂) biosynthetic genes of *Salmonella typhimurium*. *J. Bacteriol.*, 175:3303–3316, 1993.
- [245] J. Samuel, Y. Luo, P.M. Morgan, N.C. Strynadka, and M.E. Tanner. Catalysis and binding in L-ribulose-5-phosphate 4-epimerase: a comparison with L-fuculose-1-phosphate aldolase. *Biochemistry*, 40:14772–14780, 2001.
- [246] A. Schenzle, H. Lenke, J.C. Spain, and H.J. Knackmuss. 3-Hydroxylaminophenol mutase from *Ralstonia eutropha* JMP134 catalyzes a Bamberger rearrangement. *J. Bacteriol.*, 181:1444–1450, 1999.
- [247] E. Schmidt and H.-J. Knackmuss. Chemical structure and biodegradability of halogenated aromatic compounds. Conversion of chlorinated muconic acids into maleoylactic acid. *Biochem. J.*, 192:339–347, 1980.
- [248] W. Schmitz, R. Fingerhut, , and E. Purification and properties of an α -methylacyl-CoA racemase from rat liver. *Eur. J. Biochem.*, 222:313–323, 1994.
- [249] U. Schwien, E. Schmidt, H.-J. Knackmuss, and W. Reinecke. Degradation of chlorosubstituted aromatic-compounds by *Pseudomonas* sp. strain-B13 - fate of 3,5-dichlorocatechol. *Arch. Microbiol.*, 150:78–84, 1988.
- [250] B. Seckler and K. Poralla. Characterization and partial purification of squalene-hopene cyclase from *Bacillus acidocaldarius*. *Biochim. Biophys. Acta*, 881:356–363, 1986.
- [251] H.M. Seidel, S. Freeman, and J.R. Knowles. Phosphonate biosynthesis: isolation of the enzyme responsible for the formation of a carbon-phosphorus bond. *Nature*, 335:457–458, 1988.
- [252] S. Seltzer. Purification and properties of maleylacetone *cis-trans* isomerase from *Vibrio* 01. *J. Biol. Chem.*, 248:215–222, 1973.
- [253] R.-F. Shen and H.-H. Tai. Immunoaffinity purification and characterization of thromboxane synthase from porcine lung. *J. Biol. Chem.*, 261:11592–11599, 1986.
- [254] W.R. Sherman, M.A. Stewart, and M. Zinbo. Mass spectrometric study on the mechanism of D-glucose 6-phosphate-L-*myo*-inositol 1-phosphate cyclase. *J. Biol. Chem.*, 244:5703–5708, 1969.

- [255] H. Shichi and R.L. Somers. Possible involvement of retinylidene phospholipid in photoisomerization of *all-trans*-retinal to 11-*cis*-retinal. *J. Biol. Chem.*, 249:6570–6577, 1974.
- [256] Y. Shikata, T. Watanabe, T. Teramoto, A. Inoue, Y. Kawakami, Y. Nishizawa, K. Katayama, , and M. Isolation and characterization of a peptide isomerase from funnel web spider venom. *J. Biol. Chem.*, 270:16719–16723, 1995.
- [257] T. Shimizu, S. Yamamoto, and O. Hayaishi. Purification and properties of prostaglandin D synthetase from rat brain. *J. Biol. Chem.*, 254:5222–5228, 1979.
- [258] Y. Shirokane and M. Suzuki. A novel enzyme, maltose 1-epimerase from *Lactobacillus brevis* IFO 3345. *FEBS Lett.*, 367:177–179, 1995.
- [259] H. Sielaff, E. Dittmann, N. Tandeau De Marsac, C. Bouchier, H. von Döhren, T. Börner, and T. Schwecke. The *mcyF* gene of the microcystin biosynthetic gene cluster from *Microcystis aeruginosa* encodes an aspartate racemase. *Biochem. J.*, 373:909–916, 2003.
- [260] J.E. Silbert and D.H. Brown. Enzymic synthesis of uridine diphosphate glucosamine and heparin from [14C]glucosamine by a mouse mast-cell tumor. *Biochim. Biophys. Acta*, 54:590–592, 1961.
- [261] W.R. Sistrof and R.Y. Stanier. The mechanism of formation of β -keto adipic acid by bacteria. *J. Biol. Chem.*, 210:821–836, 1954.
- [262] M.W. Slein. Phosphomannose isomerase. *J. Biol. Chem.*, 186:753–761, 1950.
- [263] M.W. Slein. Xylose isomerase from *Pasteurella pestis*, strain A-1122. *J. Am. Chem. Soc.*, 77:1663–1667, 1955.
- [264] D.M. Small and N.K. Matheson. Phosphomannomutase and phosphoglucomutase in developing *Cassia corymbosa* seeds. *Phytochemistry*, 18:1147–1150, 1979.
- [265] K. Soda and T. Osumi. Crystalline amino acid racemase with low substrate specificity. *Biochem. Biophys. Res. Commun.*, 35:363–368, 1969.
- [266] F. Solano, C. Jiménez-Cervantes, J.H. Martínez-Liarte, J.C. García-Borrón, and J.A. Lozano. Molecular mechanism for catalysis by a new zinc enzyme, dopachrome tautomerase. *Biochem. J.*, 313:447–453, 1996.
- [267] R. Somack and R.N. Costilow. Purification and properties of a pyridoxal phosphate and coenzyme B₁₂ dependent D- α -ornithine 5,4-aminomutase. *Biochemistry*, 12:2597–2604, 1973.
- [268] B. Sprössler and F. Lingens. Chorismat-Mutase aus *Claviceps*. I. Eigenschaften der Chorismat-Mutase aus verschiedenen *Claviceps*-Stämmen. *Hoppe-Seyler's Z. Physiol. Chem.*, 351:448–458, 1970.
- [269] T.C. Stadtman and P. Elliott. Studies on the enzymic reduction of amino acids. II. Purification and properties of a D-proline reductase and a proline racemase from *Clostridium sticklandii*. *J. Biol. Chem.*, 228:983–997, 1957.
- [270] T.C. Stadtman and P. Renz. Anaerobic degradation of lysine. V. Some properties of the cobamide coenzyme-dependent β -lysine mutase of *Clostridium sticklandii*. *Arch. Biochem. Biophys.*, 125:226–239, 1968.
- [271] T.C. Stadtman and L. Tasi. A cobamide coenzyme dependent migration of the ϵ -amino group of D-lysine. *Biochem. Biophys. Res. Commun.*, 28:920–926, 1967.
- [272] J.R. Stern, A. del Campillo, and A.L. Lehninger. Enzymatic racemization of β -hydroxybutyryl-S-CoA and the stereospecificity of enzymes of the fatty acid cycle. *J. Am. Chem. Soc.*, 77:1073–1074, 1955.
- [273] W. Stoffel, R. Ditzer, and H. Caesar. Der Stoffwechsel der ungesättigten Fettsäuren. III. Zur β -Oxydation der Mono- und Polyenfettsäuren. Der Mechanismus der enzymatischen Reaktionen an Δ^3 *cis*-Enoyl-CoA-Verbindungen. *Hoppe-Seyler's Z. Physiol. Chem.*, 339:167–181, 1964.
- [274] W. Stoffel and W. Ecker. Δ^3 -*cis*,- Δ^2 -*trans*-Enoyl-CoA isomerase from rat liver mitochondria. *Methods Enzymol.*, 14:99–105, 1979.
- [275] W. Stoffel and M. Grol. Purification and properties of 3-*cis*-2-*trans*-enoyl-CoA isomerase (dodecenoyl-CoA Δ -isomerase) from rat liver mitochondria. *Hoppe-Seyler's Z. Physiol. Chem.*, 359:1777–1782, 1978.

- [276] P.K. Stumpf and B.L. Horecker. The rôle of xylulose 5-phosphate in xylose metabolism of *Lactobacillus pentosus*. *J. Biol. Chem.*, 218:753–768, 1956.
- [277] T.P. Sun and Y. Kamiya. The *Arabidopsis* GA1 locus encodes the cyclase *ent*-kaurene synthetase A of gibberellin biosynthesis. *Plant Cell*, 6:1509–1518, 1994.
- [278] E.W. Sutherland, M. Cohn, T. Posternak, and C.F. Cori. The mechanism of the phosphoglucomutase reaction. *J. Biol. Chem.*, 180:1285–1295, 1949.
- [279] S. Suzuki, N. Onishi, and K. Yokozeki. Purification and characterization of hydantoin racemase from *Microbacterium liquefaciens* AJ 3912. *Biosci. Biotechnol. Biochem.*, 69:530–536, 2005.
- [280] N. Takahashi, T. Hayano, and M. Suzuki. Peptidyl-prolyl *cis-trans* isomerase is the cyclosporin A-binding protein cyclophilin. *Nature*, 337:473–475, 1989.
- [281] P. Talalay and V.S. Wang. Enzymic isomerization of Δ^5 -3-ketosteroids. *Biochim. Biophys. Acta*, 18:300–301, 1955.
- [282] Y. Tanaka, S.L. Ward, and W.L. Smith. Immunochemical and kinetic evidence for two different prostaglandin *H*-prostaglandin E isomerases in sheep vesicular gland microsomes. *J. Biol. Chem.*, 262:1374–1381, 1987.
- [283] E.J. Taylor, S.J. Charnock, J. Colby, G.J. Davies, and G.W. Black. Cloning, purification and characterization of the 6-phospho-3-hexulose isomerase YckF from *Bacillus subtilis*. *Acta Crystallogr. D Biol. Crystallogr.*, 57:1138–1140, 2001.
- [284] M.B. Taylor and E. Juni. Stereoisomeric specificities of 2,3-butanediol dehydrogenase. *Biochim. Biophys. Acta*, 39:448–457, 1960.
- [285] P.L. Taylor, K.M. Blakely, G.P. de Leon, J.R. Walker, F. McArthur, E. Evdokimova, K. Zhang, M.A. Valvano, G.D. Wright, and M.S. Junop. Structure and function of sedoheptulose-7-phosphate isomerase, a critical enzyme for lipopolysaccharide biosynthesis and a target for antibiotic adjuvants. *J. Biol. Chem.*, 283:2835–2845, 2008.
- [286] H. Terada, K. Mukae, S. Hosomi, T. Mizoguchi, and K. Uehara. Characterization of an enzyme which catalyzes isomerization and epimerization of D-erythrose 4-phosphate. *Eur. J. Biochem.*, 148:345–351, 1985.
- [287] D. Thibaut, M. Couder, A. Famechon, L. Debussche, B. Cameron, J. Crouzet, , and F. The final step in the biosynthesis of hydrogenopyrnic acid is catalyzed by the cobH gene product with precorrin-8X as the substrate. *J. Bacteriol.*, 174:1043–1049, 1992.
- [288] J.B. Thoden, J. Kim, F.M. Raushel, and H.M. Holden. The catalytic mechanism of galactose mutarotase. *Protein Sci.*, 12:1051–1059, 2003.
- [289] J.B. Thoden, D.J. Timson, R.J. Reece, and H.M. Holden. Molecular structure of human galactose mutarotase. *J. Biol. Chem.*, 279:23431–23437, 2004.
- [290] L.C. Thompson, J.E. Ladner, S.G. Codreanu, J. Harp, G.L. Gilliland, and R.N. Armstrong. 2-Hydroxychromene-2-carboxylic acid isomerase: a kappa class glutathione transferase from *Pseudomonas putida*. *Biochemistry*, 46:6710–6722, 2007.
- [291] M. Tomoeda and R. Kitamura. A *cis-trans* isomerising activity of *Escherichia coli*. Isomerization from 2-(2-furyl)-3-*cis*-(5-nitro-2-furyl) acrylamide (furylfuramide) to its *trans* isomer. *Biochim. Biophys. Acta*, 480:315–325, 1977.
- [292] T. Toyomasu, H. Kawaide, A. Ishizaki, S. Shinoda, M. Otsuka, W. Mitsunashi, and T. Sassa. Cloning of a full-length cDNA encoding *ent*-kaurene synthase from *Gibberella fujikuroi*: functional analysis of a bifunctional diterpene cyclase. *Biosci. Biotechnol. Biochem.*, 64:660–664, 2000.
- [293] P.C. Trackman and R.H. Abeles. Methionine synthesis from 5'-S-methylthioadenosine. Resolution of enzyme activities and identification of 1-phospho-5-S-methylthioribulose. *J. Biol. Chem.*, 258:6717–6720, 1983.
- [294] A.G. Trejo, G.J.F. Chittenden, J.G. Buchanan, and J. Baddiley. Uridine diphosphate α -D-galactofuranose, an intermediate in the biosynthesis of galactofuranosyl residues. *Biochem. J.*, 117:637–639, 1970.
- [295] K.K. Tsuboi, J. Estrada, and P.B. Hudson. Enzymes of the human erythrocytes. IV. Phosphoglucose isomerase, purification and properties. *J. Biol. Chem.*, 231:19–29, 1958.

- [296] T.R. Tyler and J.M. Leatherwood. Epimerization of disaccharides by enzyme preparations from *Ruminococcus albus*. *Arch. Biochem. Biophys.*, 119:363–367, 1967.
- [297] V. Ullrich, L. Castle, and P. Weber. Spectral evidence for the cytochrome P_{450} nature of prostacyclin synthetase. *Biochem. Pharmacol.*, 30:2033–2036, 1981.
- [298] V. Ullrich and M. Haurand. Thromboxane synthase as a cytochrome P_{450} enzyme. *Adv. Prostaglandin Thromboxane Res.*, 11:105–110, 1983.
- [299] S. Usui and C.-A. Yu. Purification and properties of isopenicillin-N epimerase from *Streptomyces clavuligerus*. *Biochim. Biophys. Acta*, 999:78–85, 1989.
- [300] M. Uziel and D.J. Hanahan. An enzyme-catalyzed acyl migration: a lysolecithin migratase. *J. Biol. Chem.*, 226:789–798, 1957.
- [301] M.A. Valvano, P. Messner, and P. Kosma. Novel pathways for biosynthesis of nucleotide-activated glycerol-manno-heptose precursors of bacterial glycoproteins and cell surface polysaccharides. *Microbiology*, 148:1979–1989, 2002.
- [302] L. van der Drift, G.D. Vogels, and C. van der Drift. Allantoin racemase: a new enzyme from *Pseudomonas* species. *Biochim. Biophys. Acta*, 391:240–248, 1975.
- [303] L.J. van Tegelen, P.R. Moreno, A.F. Croes, R. Verpoorte, and G.J. Wullems. Purification and cDNA cloning of isochorismate synthase from elicited cell cultures of *Catharanthus roseus*. *Plant Physiol.*, 119:705–712, 1999.
- [304] A.I. Virtanen. On enzymic and chemical reactions in crushed plants. *Arch. Biochem. Biophys. Suppl.*, 1:200–208, 1962.
- [305] W.A. Volk. Purification and properties of phosphoarabinoisomerase from *Propionibacterium pentosaceum*. *J. Biol. Chem.*, 235:1550–1553, 1960.
- [306] W.M. De Vos, I. Boerrigter, R.J. Van Rooijen, B. Reiche, , and W. Characterization of the lactose-specific enzymes of the phosphotransferase system in *Lactococcus lactis*. *J. Biol. Chem.*, 265:22554–22560, 1990.
- [307] S.J. Wakil. D(-) β -Hydroxybutyryl CoA dehydrogenase. *Biochim. Biophys. Acta*, 18:314–315, 1955.
- [308] K. Watabe, T. Ishikawa, Y. Mukohara, and H. Nakamura. Purification and characterization of the hydantoin racemase of *Pseudomonas* sp. strain NS671 expressed in *Escherichia coli*. *J. Bacteriol.*, 174:7989–7995, 1992.
- [309] H. Weissbach, J. Toohey, and H.A. Barker. Isolation and properties of B_{12} coenzymes containing benzimidazole or dimethylbenzimidazole. *Proc. Natl. Acad. Sci. USA*, 45:521–528, 1959.
- [310] E.L. Westman, D.J. McNally, M. Rejzek, W.L. Miller, V.S. Kannathasan, A. Preston, D.J. Maskell, R.A. Field, J.R. Brisson, and J.S. Lam. Erratum report: Identification and biochemical characterization of two novel UDP-2,3-diacetamido-2,3-dideoxy- α -D-glucuronic acid 2-epimerases from respiratory pathogens. *Biochem. J.*, 405:625–625, 2007.
- [311] E.L. Westman, D.J. McNally, M. Rejzek, W.L. Miller, V.S. Kannathasan, A. Preston, D.J. Maskell, R.A. Field, J.R. Brisson, and J.S. Lam. Identification and biochemical characterization of two novel UDP-2,3-diacetamido-2,3-dideoxy- α -D-glucuronic acid 2-epimerases from respiratory pathogens. *Biochem. J.*, 405:123–130, 2007.
- [312] A. Wiese, M. Pietzsch, C. Syldatk, R. Mattes, and J. Altenbuchner. Hydantoin racemase from *Arthrobacter aurescens* DSM 3747: heterologous expression, purification and characterization. *J. Biotechnol.*, 80:217–230, 2000.
- [313] B.A. Wilson, S. Bantia, G.M. Salituro, A.M. Reeve, and C.A. Townsend. Cell-free biosynthesis of nocardicin A from nocardicin E and S-adenosylmethionine. . *J. Am. Chem. Soc.*, 110:8238–8239, 1988.
- [314] D.B. Wilson and D.S. Hogness. The enzymes of the galactose operon in *Escherichia coli*. I. Purification and characterization of uridine diphosphogalactose 4-epimerase. *J. Biol. Chem.*, 239:2469–2481, 1964.
- [315] D.C. Wilton, A.D. Rahimtula, and M. Akhtar. The reversibility of the Δ^8 -cholestenol- Δ^7 -cholestenol isomerase reaction in cholesterol biosynthesis. *Biochem. J.*, 114:71–73, 1969.
- [316] F.E. De Windt and D. van der Drift. Purification and some properties of hydroxypyruvate isomerase of *Bacillus fastidiosus*. *Biochim. Biophys. Acta*, 613:556–562, 1980.

- [317] M.L. Wise, M. Hamberg, and W.H. Gerwick. Biosynthesis of conjugated fatty acids by a novel isomerase from the red marine alga *Ptilota filicina*. *Biochemistry*, 33:15223–15232, 1994.
- [318] M.L. Wise, J. Rossi, and W.H. Gerwick. Binding site characterization of polyenoic fatty-acid isomerase from the marine alga *Ptilota filicina*. *Biochemistry*, 36:2985–2992, 1997.
- [319] M.L. Wise, K. Soderstrom, T.F. Murray, and W.H. Gerwick. Synthesis and cannabinoid receptor binding activity of conjugated triene anandamide, a novel eicosanoid. *Experientia*, 52:88–92, 1996.
- [320] M.J. Wolin, F.J. Simpson, and W.A. Wood. Degradation of L-arabinose by *Aerobacter aerogenes*. III. Identification and properties of L-ribulose-5-phosphate 4-epimerase. *J. Biol. Chem.*, 232:559–575, 1958.
- [321] H. Wolosker, S. Blackshaw, and S.H. Snyder. Serine racemase: a glial enzyme synthesizing D-serine to regulate glutamate-N-methyl-D-aspartate neurotransmission. *Proc. Natl. Acad. Sci. USA*, 96:13409–13414, 1999.
- [322] H. Wolosker, K.N. Sheth, M. Takahashi, J.P. Mothet, R.O. Brady, Ferris Jr., Snyder C.D., and S.H. Purification of serine racemase: biosynthesis of the neuromodulator D-serine. *Proc. Natl. Acad. Sci. USA*, 96:721–725, 1999.
- [323] W.A. Wood. Amino acid racemases. *Methods Enzymol.*, 2:212–217, 1955.
- [324] W.A. Wood and I.C. Gunsalus. D-Alanine formation: a racemase in *Streptococcus faecalis*. *J. Biol. Chem.*, 190:403–416, 1951.
- [325] T.S. Woodin and L. Nishioka. Evidence for three isozymes of chorismate mutase in alfalfa. *Biochim. Biophys. Acta*, 309:211–223, 1973.
- [326] B. Wurster and B. Hess. Glucose-6-phosphate-1-epimerase from baker's yeast. A new enzyme. *FEBS Lett.*, 23:341–348, 1972.
- [327] C.J. Xu, D.M. Chen, and S.L. Zhang. [Molecular cloning of full length capsanthin/capsorubin synthase homologous gene from orange (*Citrus sinensis*)]. *Shi Yan Sheng Wu Xue Bao*, 34:147–150, 2001.
- [328] M. Xu, M.L. Hillwig, S. Priscic, R.M. Coates, and R.J. Peters. Functional identification of rice *syn*-copalyl diphosphate synthase and its role in initiating biosynthesis of diterpenoid phytoalexin/allelopathic natural products. *Plant J.*, 39:309–318, 2004.
- [329] M. Yamada and K. Kurahashi. Further purification and properties of adenosine triphosphate-dependent phenylalanine racemase of *Bacillus brevis* Nagano. *J. Biochem. (Tokyo)*, 66:529–540, 1969.
- [330] K. Yamanaka. Purification, crystallization and properties of the D-xylose isomerase from *Lactobacillus brevis*. *Biochim. Biophys. Acta*, 151:670–680, 1968.
- [331] T. Yamashita, M. Ashiuchi, K. Ohnishi, S. Kato, S. Nagata, and H. Misono. Molecular identification of monomeric aspartate racemase from *Bifidobacterium bifidum*. *Eur. J. Biochem.*, 271:4798–4803, 2004.
- [332] T. Yamauchi, S.Y. Choi, H. Okada, M. Yohda, H. Kumagai, N. Esaki, and K. Soda. Properties of aspartate racemase, a pyridoxal 5'-phosphate-independent amino acid racemase. *J. Biol. Chem.*, 267:18361–18364, 1992.
- [333] W.K. Yeh, S.K. Ghag, and S.W. Queener. Enzymes for epimerization of isopenicillin N, ring expansion of penicillin N, and 3'-hydroxylation of deacetoxycephalosporin C. Function, evolution, refolding, and enzyme engineering. *Ann. N.Y. Acad. Sci.*, 672:396–408, 1992.
- [334] W.S. Yew and J.A. Gerlt. Utilization of L-ascorbate by *Escherichia coli* K-12: assignments of functions to products of the *yjf-sga* and *yia-sgb* operons. *J. Bacteriol.*, 184:302–306, 2002.
- [335] T. Yorifuji, K. Ogata, and K. Soda. Crystalline arginine racemase. *Biochem. Biophys. Res. Commun.*, 34:760–764, 1969.
- [336] I.G. Young and F. Gibson. Regulation of the enzymes involved in the biosynthesis of 2,3-dihydroxybenzoic acid in *Aerobacter aerogenes* and *Escherichia coli*. *Biochim. Biophys. Acta*, 177:401–411, 1969.
- [337] S. Yu and R. Fiskesund. The anhydrofructose pathway and its possible role in stress response and signaling. *Biochim. Biophys. Acta*, 1760:1314–1322, 2006.

- [338] S. Yu, C. Refdahl, and I. Lundt. Enzymatic description of the anhydrofructose pathway of glycogen degradation; I. Identification and purification of anhydrofructose dehydratase, ascopyrone tautomerase and α -1,4-glucan lyase in the fungus *Anthracobia melaloma*. *Biochim. Biophys. Acta*, 1672:120–129, 2004.
- [339] H. Yurimoto, N. Kato, and Y. Sakai. Assimilation, dissimilation, and detoxification of formaldehyde, a central metabolic intermediate of methylotrophic metabolism. *Chem. Rec.*, 5:367–375, 2005.
- [340] V. Zappia and H.A. Barker. Studies on lysine-2,3-aminomutase. Subunit structure and sulfhydryl groups. *Biochim. Biophys. Acta*, 207:505–513, 1970.
- [341] W. Zheng, M.L. Wise, A. Wyrick, J.G. Metz, L. Yuan, and W.H. Gerwick. Polyenoic fatty-acid isomerase from the marine red alga *Ptilota filicina*: protein characterization and functional expression of the cloned cDNA. *Arch. Biochem. Biophys.*, 401:11–20, 2002.

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